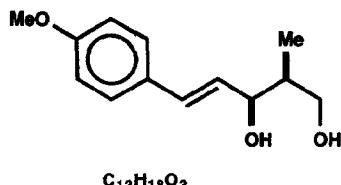


STEREOCHEMISTRY ABSTRACTS

H. Akita, I. Umezawa, M. Nozawa, S. Nagumo

Tetrahedron: Asymmetry 1993, 4, 757



3-Hydroxy-5-(4'-methoxyphenyl)-  
2-methyl-(4E)-penten-1-ol

E.e. = >99% [by chiral HPLC]

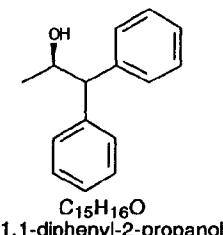
$[\alpha]_D^{25} +17.4$  ( $c=1.00$ ,  $CHCl_3$ )

Source of chirality: immobilized lipase  
catalysed resolution

Absolute configuration 2S, 3S

L.R. Randrianasolo-Rakotozafy, R. Azerad, F. Dumas,  
D. Potin and J. d'Angelo

Tetrahedron: Asymmetry 1993, 4, 761



1,1-diphenyl-2-propanol

E.e. = 96.5 % [by GC analysis of the corresponding  
(S)-O-acetylactyl ester]

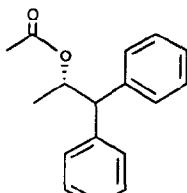
$[\alpha]_D^{25} = +44$  ( $c = 0.55$ , MeOH)

Source of chirality: enzymatic hydrolysis of the  
acetate

Absolute configuration: (R)- (assigned by optical  
rotation)

L.R. Randrianasolo-Rakotozafy, R. Azerad, F. Dumas,  
D. Potin and J. d'Angelo

Tetrahedron: Asymmetry 1993, 4, 761



$C_{17}H_{18}O_2$   
2-acetoxy-1,1-diphenylpropane

E.e. = 98 % [by GC analysis after saponification and  
derivatization to the corresponding (S)-O-acetylactyl  
ester]

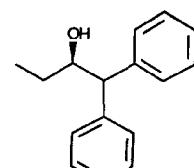
$[\alpha]_D^{25} = -50.7$  ( $c = 3.25$ , MeOH)

Source of chirality: enzymatic hydrolysis

Absolute configuration: (S)- (assigned by optical  
rotation after saponification)

L.R. Randrianasolo-Rakotozafy, R. Azerad, F. Dumas,  
D. Potin and J. d'Angelo

Tetrahedron: Asymmetry 1993, 4, 761



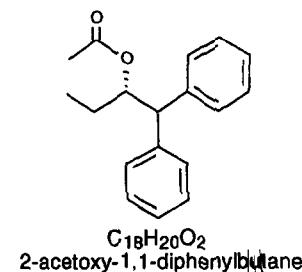
$C_{16}H_{18}O$   
1,1-diphenyl-2-butanol

E.e. = 94 % [by GC analysis of the corresponding  
(S)-O-acetylactyl ester]

$[\alpha]_D^{25} = +35.4$  ( $c = 3.2$ , MeOH)

Source of chirality: enzymatic hydrolysis of the  
acetate

Absolute configuration: (R)- (assigned by optical  
rotation)

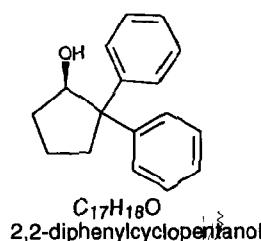


E.e. = 84 % [by GC analysis after saponification and derivatization to the corresponding (S)-O-acetylactyl ester]

$[\alpha]_D^{25} = -9.5$  ( $c = 3.25$ , MeOH)

Source of chirality: enzymatic hydrolysis

Absolute configuration: (S)- (assigned by optical rotation after saponification)

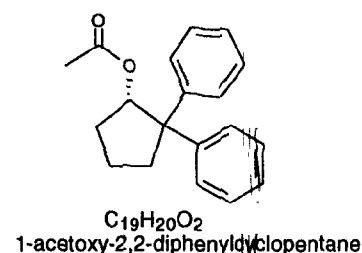


E.e. = 96.5% [by GC analysis of the corresponding (S)-O-acetylactyl ester]

$[\alpha]_D^{25} = -109.4$  ( $c \approx 1.1$ , EtOH)

Source of chirality: enzymatic hydrolysis of the acetate

Absolute configuration: (R)- (assigned by optical rotation)

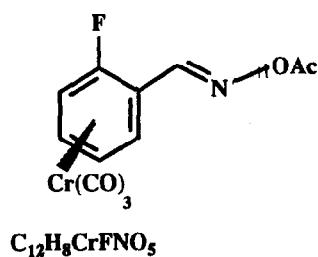


E.e. = 92 % [by GC analysis after saponification and derivatization to the corresponding (S)-O-acetylactyl ester]

$[\alpha]_D^{25} = +121.8$  ( $c \approx 0.62$ , EtOH)

Source of chirality: enzymatic hydrolysis

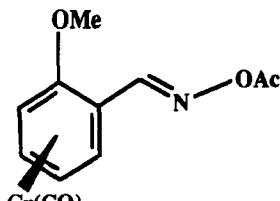
Absolute configuration: (S)- (assigned by optical rotation after saponification)



E.e. = 72 % by chiral HPLC with a Chiracel OB column

Source of chirality : *Humicola lanuginosa* lipase

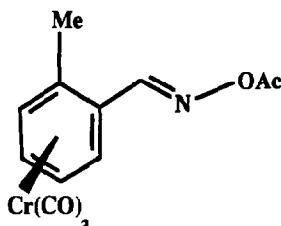
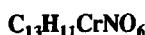
Absolute configuration : 1S



E.e. = 76 % by chiral HPLC with a Chiralcel OD column

Source of chirality : *Pseudomonas cepacia* lipase

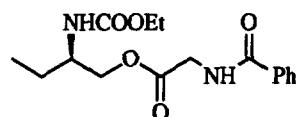
Absolute configuration : 1S



E.e. = 98 % by chiral HPLC with a Chiralcel OJ column

Source of chirality : *Pseudomonas cepacia* lipase

Absolute configuration : 1S

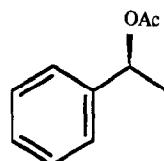
[ $\alpha$ ] $^{25}_D$  = +21.5 ( 6, EtOH )

Source of chirality: (R)-2-amino-1-butanol

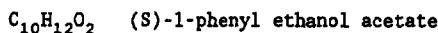
Absolute configuration: R



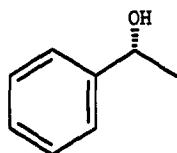
2-[(N-Ethoxycarbonyl)amino]butyl hippurate



E.e. = 99 % by chiral HPLC of the alcohol

[ $\alpha$ ] $^{25}_D$  = -106.0 ( c=1, ether )Source of chirality : Native/modified Lipase PS  
catalyzed resolution

(S)-1-phenyl ethanol acetate      Absolute configuration: (S)



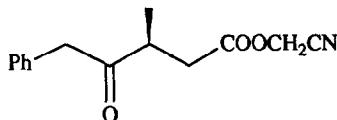
$C_8H_{10}O$  (R)-1-phenyl ethanol

E.e. = 99 % by chiral HPLC

$[\alpha]_D^{25} = +41.0$  (neat)

Source of chirality : Native/modified Lipase PS catalyzed resolution

Absolute configuration: (R)



$C_{14}H_{15}O_3N$

Cyanomethyl 3-methyl-4-oxo-5-phenylpentanoate

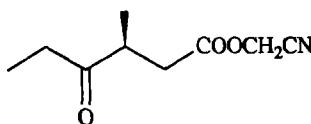
E.e. > 95% [by nmr with Eu(hfc)<sub>3</sub>]

$[\alpha]_D^{20} = +12.9$  (c 1.6, THF)

Source of chirality: enzymatic resolution

Absolute configuration 3R

(assigned by chemical correlation)



$C_9H_{13}O_3N$

Cyanomethyl 3-methyl-4-oxohexanoate

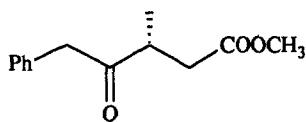
E.e.=93% [by nmr with Eu(hfc)<sub>3</sub>]

$[\alpha]_D^{20} = +21.8$  (c = 1.85, THF)

Source of chirality: enzymatic resolution

Absolute configuration 3R

(assigned by chemical correlation)



$C_{13}H_{16}O_3$

Methyl 3-methyl-4-oxo-5-phenylpentanoate

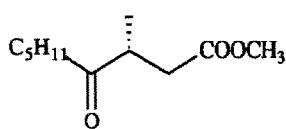
E.e.= 85% [by nmr with Eu(hfc)<sub>3</sub>]

$[\alpha]_D^{20} = -10.9$  (c 3.0, THF)

Source of chirality: enzymatic resolution

Absolute configuration 3S

(assigned by chemical correlation)

 $C_{11}H_{20}O_3$ 

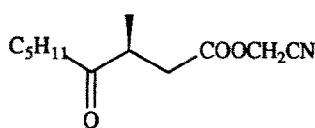
Methyl 3-methyl-4-oxononanoate

E.e.=85% [by nmr with Eu(hfc)<sub>3</sub>] $[\alpha]_D^{20} = -30.1$  (c 2.9, THF)

Source of chirality: enzymatic resolution

Absolute configuration 3S

(assigned by chemical correlation)

 $C_{12}H_{19}O_3N$ 

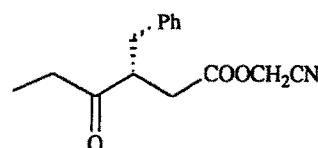
Cyanomethyl 3-methyl-4-oxononanoate

E.e.=92% [by nmr with Eu(hfc)<sub>3</sub>] $[\alpha]_D^{20} = +22.6$  (c 3.7, THF)

Source of chirality: enzymatic resolution

Absolute configuration 3R

(assigned by chemical correlation)

 $C_{15}H_{17}O_3N$ 

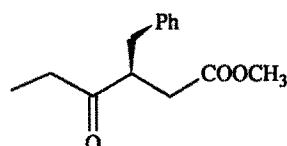
Cyanomethyl 3-benzyl-4-oxo-5-hexanoate

E.e.> 98% [by nmr with Eu(hfc)<sub>3</sub>] $[\alpha]_D^{20} = -72.8$  (c 6.5, THF)

Source of chirality: enzymatic resolution

Absolute configuration 3S

(assigned by chemical correlation)

 $C_{14}H_{18}O_3$ 

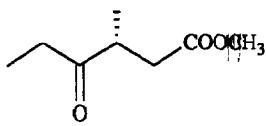
Methyl 3-benzyl-4-oxo-5-hexanoate

E.e.= 81% [by nmr with Eu(hfc)<sub>3</sub>] $[\alpha]_D^{20} = +86.6$  (c 1.4, THF)

Source of chirality: enzymatic resolution

Absolute configuration 3R

(assigned by chemical correlation)

 $C_8H_{14}O_3$ 

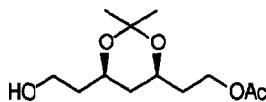
Methyl 3-methyl-4-oxohexanoate

E.e.=81% [by nmr with  $\text{Eu}(\text{hfc})_3$ ] $[\alpha]_D^{20} = -34.8$  (c 1.6, THF)

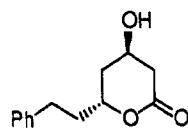
Source of chirality: enzymatic resolution

Absolute configuration 3S

(assigned by chemical correlation)

E.e.=>98% by  $^1\text{H-NMR}$  with  $\text{Eu}(\text{hfc})_3$  $[\alpha]_D = -12.8$  (c= 2.8,  $\text{CHCl}_3$ )source of chirality: enzymatic hydrolysis  
of meso diacetate

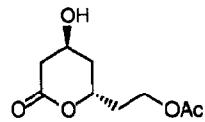
(3S,5R)-7-hydroxy-3,5-O-isopropylidene-1-acetoxy heptane



E.e.= &gt; 98%

 $[\alpha]_D = +48$  (c= 2.2,  $\text{CHCl}_3$ )source of chirality: (3S,5R)-7-hydroxy-  
3,5-O-isopropylidene-1-acetoxy heptane

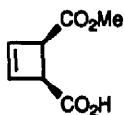
(3R,5R)-3-hydroxy-1-phenylheptan-5-olide



E.e. &gt;98%

 $[\alpha]_D = -16$  (c= 0.75,  $\text{CHCl}_3$ )source of chirality: (3S,5R)-7-hydroxy-  
3,5-O-isopropylidene-1-acetoxy heptane

(3S,5S)-3-hydroxy-1-acetoxyheptan-5-olide



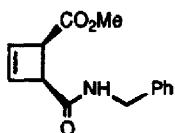
Methyl hydrogen (1R, 2S)-  
(+)-3-cyclobutene-1,2-dicarboxylate

E.e = 86% (by nmr as the salt with (S)-(−)- $\alpha$ -methylbenzylamine  
 $[\alpha]_D^{20} = 6.9^\circ$  (c 2.5 in  $\text{CHCl}_3$ )

Source of chirality: kinetic resolution using porcine liver esterase

Absolute configuration 1R, 2S

(from comparison of sign of optical rotation with literature value)



Methyl (1R,  
2S)-(+)-2-benzylcarbamoyl-  
3-cyclobutene carboxylate

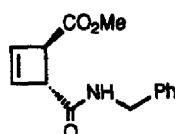
E.e = 100% (assumed from crystallisation to constant optical rotation  
of material of 86% ee).

$[\alpha]_D^{20} = 11.7^\circ$  (c 1.05 in  $\text{CHCl}_3$ )

Source of chirality: kinetic resolution using porcine liver esterase

Absolute configuration 1R, 2S

(from method of synthesis)



Methyl (1R,  
2R)-(+)-2-benzylcarbamoyl-  
3-cyclobutene carboxylate

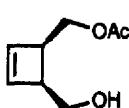
E.e = 100% (assumed from crystallisation to constant optical rotation  
of material of 86% ee).

$[\alpha]_D^{20} = -255^\circ$  (c 1.0 in  $\text{CHCl}_3$ )

Source of chirality: kinetic resolution using porcine liver esterase

Absolute configuration 1R, 2R

(from method of synthesis)



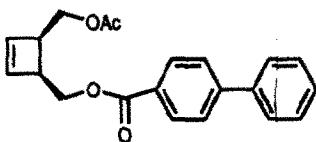
((1R,4S)-4-Hydroxymethyl-  
2-cyclobutenyl)methyl  
ethanoate

E.e = >97% (by nmr with (S)-(+)-2,2,2-trifluoro-1-(9-anthryl)ethanol).  
 $[\alpha]_D^{20} = 6.2^\circ$  (c 2.0 in  $\text{CHCl}_3$ )

Source of chirality: kinetic resolution using *Pseudomonas* sp. lipase

Absolute configuration 1R, 4S

(from method of synthesis)



((1R,4S)-4-(4-Phenylphenyl)methoxyxymethyl-2-cyclobutenyl)methyl ethanoate

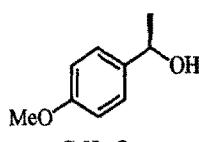
E.e. = 100% (assumed from crystallisation to constant optical rotation of material of >97% ee).

$[\alpha]_D^{20} = -11.1^\circ$  (c 1.0 in CHCl<sub>3</sub>)

Source of chirality: kinetic resolution using *Pseudomonas* sp. lipase

Absolute configuration 1R, 4S

(from method of synthesis)



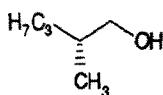
1-(4-methoxyphenyl)ethanol

e.e. >95% by Mosher's ester and Eu(hfc)<sub>3</sub>

$[\alpha]_D^{22} = +58.6$  (c 1.0, CHCl<sub>3</sub>)

Source of chirality: Lipase mediated double kinetic resolution

Absolute configuration: R (cf JACS 1989, 111, 3426).



2-Methyl-1-pentanol

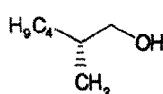
E.e. = 98% [by gas chromatography on  $\beta$ -cyclodextrin phase]

$[\alpha]_D^{20} = +12.10$  (neat)

Source of chirality: Lipase-catalyzed kinetic resolution

Absolute configuration 2R

(assigned by comparison of known optical rotation value)



2-Methyl-1-hexanol

E.e. = 99% [by gas chromatography on  $\beta$ -cyclodextrin phase]

$[\alpha]_D^{20} = +14.22$  (c=6.96, diethyl ether)

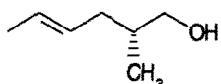
Source of chirality: Lipase-catalyzed kinetic resolution

Absolute configuration 2R

(assigned by comparison of known optical rotation value)



S. Barth and F. Effenberger\*



2-Methyl-4-hexen-1-ol

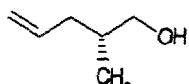
E.e. = 96.1% [by gas chromatography on  $\beta$ -cyclodextrin phase] $[\alpha]_D^{20} = +2.67$  ( $c=6.32$ ,  $\text{CH}_2\text{Cl}_2$ )

Source of chirality: Lipase-catalyzed kinetic resolution

Absolute configuration 2R

(assigned by comparison of known optical rotation value)

S. Barth and F. Effenberger\*



2-Methyl-4-penten-1-ol

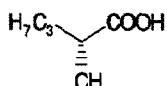
E.e. = 97.3% [by gas chromatography on  $\beta$ -cyclodextrin phase] $[\alpha]_D^{20} = +2.64$  (neat)

Source of chirality: Lipase-catalyzed kinetic resolution

Absolute configuration 2R

(assigned by comparison of known optical rotation value)

S. Barth and F. Effenberger\*



2-Methylpentanoic acid

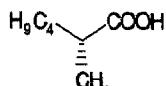
E.e. = 94.8% [by gas chromatography on  $\beta$ -cyclodextrin phase] $[\alpha]_D^{20} = -16.55$  ( $c=4.20$ ,  $\text{CHCl}_3$ )

Source of chirality: Oxidation of the optically pure alcohol (94.6%ee)

Absolute configuration 2R

(assigned by comparison of known optical rotation value)

S. Barth and F. Effenberger\*



2-Methylhexanoic acid

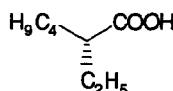
E.e. = 96.4% [by gas chromatography on  $\beta$ -cyclodextrin phase] $[\alpha]_D^{20} = -18.05$  ( $c=5.12$ ,  $\text{CHCl}_3$ )

Source of chirality: Oxidation of the optically pure alcohol (96.7%ee)

Absolute configuration 2R

(assigned by comparison of known optical rotation value)

S. Barth and F. Effenberger\*



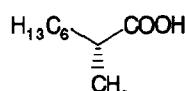
2-Ethylhexanoic acid

E.e. = 92.4% [by gas chromatography on  $\beta$ -cyclodextrin phase]  
 $[\alpha]_D^{20} = -7.43$  ( $c=3.62$ ,  $\text{CHCl}_3$ )

Source of chirality: Oxidation of the optically pure alcohol (93.5%ee)

Absolute configuration 2R  
 (assigned by comparison of known optical rotation value)

S. Barth and F. Effenberger\*



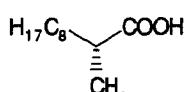
2-Methyloctanoic acid

E.e. = 93.7% [by gas chromatography on  $\beta$ -cyclodextrin phase]  
 $[\alpha]_D^{20} = -15.60$  ( $c=4.14$ ,  $\text{CHCl}_3$ )

Source of chirality: Oxidation of the optically pure alcohol (94.1%ee)

Absolute configuration 2R  
 (assigned by comparison of known optical rotation value)

S. Barth and F. Effenberger\*



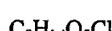
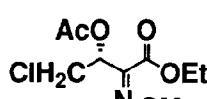
2-Methyldecanoic acid

E.e. = 96.5% [by gas chromatography on  $\beta$ -cyclodextrin phase]  
 $[\alpha]_D^{20} = -15.91$  ( $c=3.22$ ,  $\text{CHCl}_3$ )

Source of chirality: Oxidation of the optically pure alcohol (97.5%ee)

Absolute configuration 2R  
 (assigned by comparison of known optical rotation value)

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,  
 and Tamotsu Fujisawa\*



Ethyl (R)-3-Acetoxy-4-chloro-2-methoxyiminobutyrate

ee = 96% [determined by  $^1\text{H}$  NMR and GLC analysis of  
 the corresponding MTPA ester]

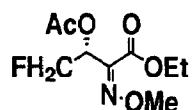
$[\alpha]_D^{23} = -59.8$  ( $c 0.98$ ,  $\text{MeOH}$ )

Source of chirality: Optical resolution by lipase

Absolute configuration: R (assigned after converting into the known ethyl  
 $(2R,3R)$ -2-amino-4-fluoro-3-hydroxybutyrate)

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,

and Tamotsu Fujisawa\*

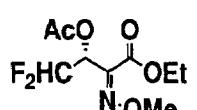
 $\text{C}_9\text{H}_{14}\text{O}_5\text{F}$ Ethyl (*R*)-3-Acetoxy-4-fluoro-2-methoxyiminobutyrateee = >98% [determined by  $^1\text{H}$  NMR and GLC analysis of the corresponding MTPA ester] $[\alpha]_D^{23} -62.8$  (c 1.24, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: *R* (assigned after converting into the known ethyl (*R*,*S*)-2-amino-4-fluoro-3-hydroxybutyrate)

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,

and Tamotsu Fujisawa\*

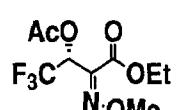
 $\text{C}_9\text{H}_{13}\text{O}_5\text{F}_2$ Ethyl (*R*)-3-Acetoxy-4,4-difluoro-2-methoxyiminobutyrateee = >98% [determined by  $^1\text{H}$  NMR and GLC analysis of the corresponding MTPA ester] $[\alpha]_D^{23} -25.5$  (c 0.22, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: *R* (assigned after converting into the known ethyl (*R*,*S*)-2-amino-4,4-difluoro-3-hydroxybutyrate)

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,

and Tamotsu Fujisawa\*

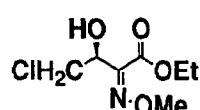
 $\text{C}_9\text{H}_{12}\text{O}_5\text{F}_3$ Ethyl (*R*)-3-Acetoxy-4,4,4-trifluoro-2-methoxyiminobutyrateee = 90% [determined by  $^1\text{H}$  NMR and GLC analysis of the corresponding MTPA ester] $[\alpha]_D^{23} +9.20$  (c 0.94, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: *R* (assigned after converting into the known ethyl (*R*,*S*)-2-amino-4,4,4-trifluoro-3-hydroxybutyrate)

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,

and Tamotsu Fujisawa\*

 $\text{C}_9\text{H}_{14}\text{O}_4\text{Cl}$ Ethyl (*S*)-4-Chloro-3-hydroxy-2-methoxyiminobutyrateee = >98% [determined by  $^1\text{H}$  NMR and GLC analysis of the corresponding MTPA ester] $[\alpha]_D^{23} +17.3$  (c 0.82, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: *S* (assigned after converting into the known ethyl (*S*,*S*)-2-amino-4-fluoro-3-hydroxybutyrate)

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,  
and Tamotsu Fujisawa\*

ee = >98% [determined by  $^1\text{H}$  NMR and GLC analysis of  
the corresponding MTPA ester]

$[\alpha]_D^{23} +19.4$  (c 1.04, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: *S*(assigned after converting into the known ethyl (2*S*,3*S*)-  
2-amino-4-fluoro-3-hydroxybutyrate)

Ethyl (*S*)-4-Fluoro-3-hydroxy-2-methoxyiminobutyrate

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,  
and Tamotsu Fujisawa\*

ee = >98% [determined by  $^1\text{H}$  NMR and GLC analysis of  
the corresponding MTPA ester]

$[\alpha]_D^{23} +8.89$  (c 0.40, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: *S* (assigned after converting into the known ethyl  
(2*S*,3*S*)-2-amino-4,4-difluoro-3-hydroxybutyrate)

Ethyl (*S*)-4,4-Difluoro-3-hydroxy-2-methoxyiminobutyrate

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,  
and Tamotsu Fujisawa\*

ee = 82% [determined by  $^1\text{H}$  NMR and GLC analysis of  
the corresponding MTPA ester]

$[\alpha]_D^{23} -64.0$  (c 0.72, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: *S* (assigned after converting into the known ethyl  
(2*S*,3*S*)-2-amino-4,4,4-trifluoro-3-hydroxybutyrate)

Ethyl (*S*)-4,4,4-Trifluoro-3-hydroxy-2-methoxyiminobutyrate

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,  
and Tamotsu Fujisawa\*

ee = >98% [determined by  $^1\text{H}$  NMR and GLC analysis of  
the corresponding bis-MTPA derivative]

$[\alpha]_D^{23} -5.83$  (c 0.24, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: (2*S*,3*S*) (assigned after converting into the known  
(2*S*,3*S*)-4-fluorothreonine)

Ethyl (2*S*,3*S*)-2-Amino-4-fluoro-3-hydroxybutyrate

Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,  
and Tamotsu Fujisawa\*

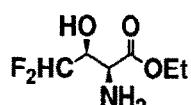
ee = >98% [determined by  $^1\text{H}$  NMR and GLC analysis of  
the corresponding bis-MTPA derivative]

$[\alpha]_D^{23} -15.0$  (c 0.08, MeOH)

Source of chirality: Optical resolution by lipase

Absolute configuration: (2S,3S) (assigned after converting into the known  
(2S,3S)-4,4-difluorothreonine)

Ethyl (2S,3S)-2-Amino-4,4-difluoro-3-hydroxybutyrate



C<sub>6</sub>H<sub>11</sub>O<sub>3</sub>F<sub>2</sub>

Ethyl (2S,3S)-2-Amino-4,4-difluoro-3-hydroxybutyrate

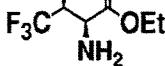
Makoto Shimizu, Tetsuya Yokota, Kouichi Fujimori,  
and Tamotsu Fujisawa\*

ee = 84% [determined by  $^1\text{H}$  NMR and GLC analysis of  
the corresponding bis-MTPA derivative]

$[\alpha]_D^{23} -8.30$  (c 0.24, MeOH)

Source of chirality: Optical resolution by lipase

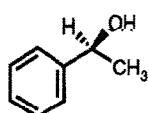
Absolute configuration: (2S,3S) (assigned after converting into the known  
(2S,3S)-4,4,4-difluorothreonine)



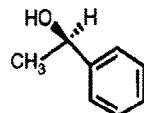
C<sub>6</sub>H<sub>10</sub>O<sub>3</sub>F<sub>3</sub>

Ethyl (2S,3S)-2-Amino-4,4,4-trifluoro-3-hydroxybutyrate

A. L. Gutman,\* D. Brenner, and A. Boltanski



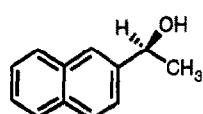
S-(-)-sec-phenethyl alcohol  
ee>99.6%



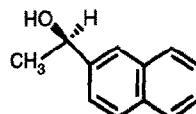
R-(+)-sec-phenethyl alcohol  
ee=98%

Source of chirality:  
enzymatic resolution  
ee determined by chiral HPLC

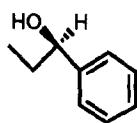
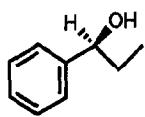
A. L. Gutman,\* D. Brenner, and A. Boltanski



S-(-)-1-(2-naphthyl)ethanol  
ee=99.8%



Source of chirality:  
enzymatic resolution  
ee determined by chiral HPLC

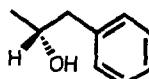
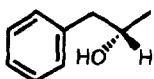


S-(-)-1-phenyl-1-propanol R-(+)-1-phenyl-1-propanol

ee=97%

ee=94%

Source of chirality:  
enzymatic resolution  
ee determined by chiral HPLC

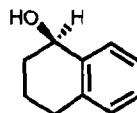
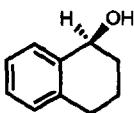


S-(+)-1-phenyl-2-propanol R-(-)-1-phenyl-2-propanol

ee=99.6%

ee=99%

Source of chirality:  
enzymatic resolution  
ee determined by chiral HPLC

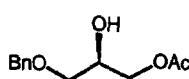


S-(+)-1,2,3,4-tetrahydro-1-naphthol R-(-)-1,2,3,4-tetrahydro-1-naphthol

ee=98.6%

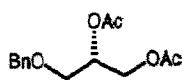
ee=94.3%

Source of chirality:  
enzymatic resolution  
ee determined by chiral HPLC

E.e. >95% [by <sup>1</sup>H-NMR in the presence of Eu(hfc)<sub>3</sub>][ $\alpha$ ]<sub>D</sub><sup>25</sup> = -4.1 (CHCl<sub>3</sub>, c = 1.04) $C_{12}H_{16}O_4$ 

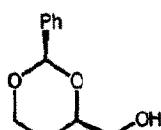
Source of chirality: lipase catalyzed transesterification.

(R)-1-O-Acetyl-3-O-benzylpropane-1,2,3-triol

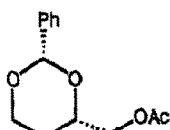
E.e. >95% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25} = +14.0$  (CHCl<sub>3</sub>, c= 0.5)C<sub>14</sub>H<sub>18</sub>O<sub>5</sub>

Source of chirality: lipase catalyzed transesterification.

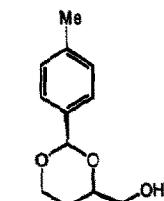
(S)-1,2-Di-O-acetyl-3-O-benzylpropane-1,2,3-triol

E.e. > 98% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25} = -10.0$  (CHCl<sub>3</sub>, c= 1.18)

Source of chirality: lipase catalyzed transesterification.

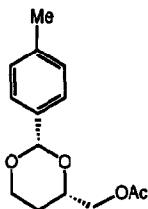
C<sub>11</sub>H<sub>14</sub>O<sub>3</sub>  
(R,R)-4-Hydroxymethyl-2-phenyl-1,3-dioxaneE.e. > 98% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25} = +27.1$  (CHCl<sub>3</sub>, c= 1.2)

Source of chirality: lipase catalyzed transesterification.

C<sub>13</sub>H<sub>16</sub>O<sub>4</sub>  
(S,S)-4-Acetoxymethyl-2-phenyl-1,3-dioxaneE.e. > 98% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25} = -8.0$  (CHCl<sub>3</sub>, c= 0.96)

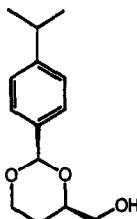
Source of chirality: lipase catalyzed transesterification.

C<sub>12</sub>H<sub>16</sub>O<sub>3</sub>  
(R,R)-4-Hydroxymethyl-2-(4-methylphenyl)-1,3-dioxane

E.e.= 84% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25}= +25.2$  (CHCl<sub>3</sub>, c= 1.05)

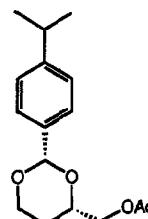
Source of chirality: lipase catalyzed transesterification.

$\text{C}_{14}\text{H}_{18}\text{O}_4$   
(S,S)-4-Acetoxyethyl-2-(4-methylphenyl)-1,3-dioxane

E.e. > 94% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25}= -11.1$  (CHCl<sub>3</sub>, c= 1.5)

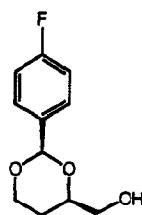
Source of chirality: lipase catalyzed transesterification.

$\text{C}_{14}\text{H}_{20}\text{O}_3$   
(R,R)-4-Hydroxymethyl-2-(4-isopropylphenyl)-1,3-dioxane

E.e.= 82% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25}= +20.9$  (CHCl<sub>3</sub>, c= 0.6)

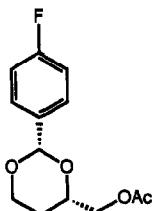
Source of chirality: lipase catalyzed transesterification.

$\text{C}_{16}\text{H}_{22}\text{O}_4$   
(S,S)-4-Acetoxyethyl-2-(4-isopropylphenyl)-1,3-dioxane

E.e.= 90% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25}= -10.9$  (CHCl<sub>3</sub>, c= 0.7)

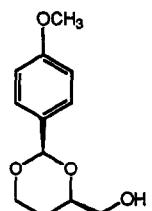
Source of chirality: lipase catalyzed transesterification.

$\text{C}_{11}\text{H}_{13}\text{FO}_3$   
(R,R)-2-(4-Fluorophenyl)-4-hydroxymethyl-1,3-dioxane

E.e.= 87% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25} = +24.1$  (CHCl<sub>3</sub>, c= 1.65)

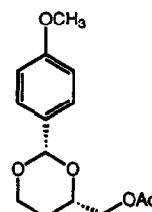
Source of chirality: lipase catalyzed transesterification.

$\text{C}_{13}\text{H}_{15}\text{FO}_4$   
(S,S)-4-Acetoxyethyl-2-(4-fluorophenyl)-1,3-dioxane

E.e.> 98% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25} = -10.3$  (CHCl<sub>3</sub>, c= 1.45)

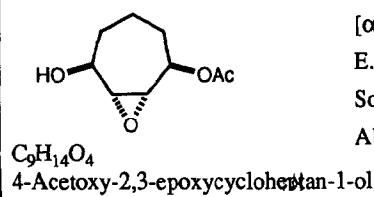
Source of chirality: lipase catalyzed transesterification.

$\text{C}_{12}\text{H}_{16}\text{O}_4$   
(R,R)-4-Hydroxymethyl-2-(4-methoxyphenyl)-1,3-dioxane

E.e.= 90% [by  $^1\text{H}$ -NMR in the presence of Eu(hfc)<sub>3</sub>] $[\alpha]_D^{25} = +25.1$  (CHCl<sub>3</sub>, c= 0.77)

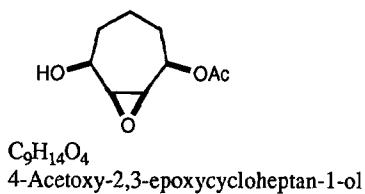
Source of chirality: lipase catalyzed transesterification.

$\text{C}_{14}\text{H}_{18}\text{O}_5$   
(S,S)-4-Acetoxyethyl-2-(4-methoxyphenyl)-1,3-dioxane

 $[\alpha]_D^{25} = +35.6$  (c 1.50, CHCl<sub>3</sub>)

E.e. = &gt;98% [optical rotation comparison to enantiomer; chemical correlation]

Source of chirality: Enzymatic asymmetric resolution of *meso*-diacetateAbsolute configuration: 1*S*, 2*R*, 3*S*, 4*R*

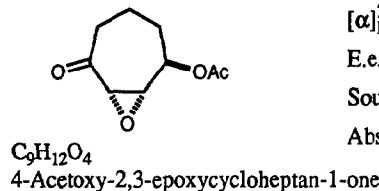


$[\alpha]_D^{25} = -12.6$  (*c* 1.05, CHCl<sub>3</sub>)

E.e. = 84% [by <sup>19</sup>F NMR of (*R*)-MTPA ester]

Source of chirality: Enzymatic asymmetrication of *meso*-diol

Absolute configuration: 1*S*, 2*S*, 3*R*, 4*R*

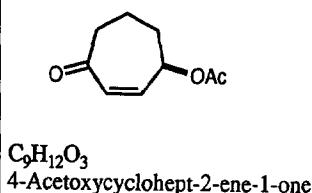


$[\alpha]_D^{25} = +26.2$  (*c* 0.69, CHCl<sub>3</sub>)

E.e. = >98% [derived from enantiomerically pure alcohol]

Source of chirality: Enzymatic asymmetrication of *meso*-diacetate

Absolute configuration: 2*S*, 3*S*, 4*R*

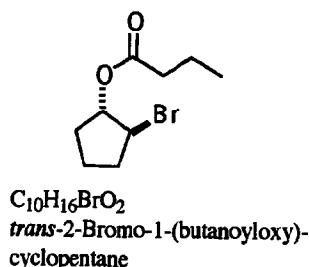


$[\alpha]_D^{25} = +106.9$  (*c* 1.00, CHCl<sub>3</sub>)

E.e. = >98% [derived from enantiomerically pure alcohol, chemical correlation]

Source of chirality: Enzymatic asymmetrication of *meso*-diacetate

Absolute configuration: 4*R*



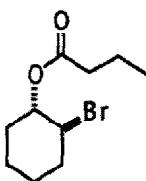
E.e. >98% [by <sup>1</sup>H-NMR in the presence of Eu(hfc)<sub>3</sub>]

$[\alpha]_D = +75.6$  (*c* 10.7, CH<sub>2</sub>Cl<sub>2</sub>)

Source of chirality: enzyme-catalyzed resolution

Absolute configuration: 1*S*, 2*S*

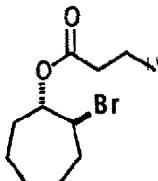
(assigned by conversion to (S)-2-cyclopenten-1-ol)

 $C_{10}H_{16}BrO_2$ *trans*-2-Bromo-1-(butanoyloxy)-cyclohexane

E.e. >98% [by  $^1\text{H}$ -NMR in the presence of  $\text{Eu}(\text{hfc})_3$ ]  
 $[\alpha]_D = +45.3$  (*c* 10.1,  $\text{CH}_2\text{Cl}_2$ ), lit.\*  $[\alpha]_D = +43.6$  (*c* 2,  $\text{CH}_2\text{Cl}_2$ )

Source of chirality: enzyme-catalyzed resolution

Absolute configuration: 1S,2S

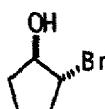
\*Hönig, H.; Seufert-Wasserthal, P. *Synthesis*, 1990, 12, 1137-1140. $C_{10}H_{16}BrO_2$ *trans*-2-Bromo-1-(butanoyloxy)-cycloheptane

E.e. >98% [by  $^1\text{H}$ -NMR in the presence of  $\text{Eu}(\text{hfc})_3$ ]  
 $[\alpha]_D = +45.3$  (*c* 10.9,  $\text{CH}_2\text{Cl}_2$ )

Source of chirality: enzyme-catalyzed resolution

Absolute configuration: 1S,2S

(assigned by conversion to (S)-2-cyclohepten-1-ol)

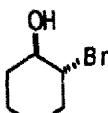
 $C_5H_9BrO$ *trans*-2-Bromo-1-cyclopentanol

E.e. = 84% [by  $^1\text{H}$ -NMR of the acetate ester in the presence of  $\text{Eu}(\text{hfc})_3$ ]  
 $[\alpha]_D = -32.1$  (*c* 12.2,  $\text{CH}_2\text{Cl}_2$ )

Source of chirality: enzyme-catalyzed resolution

Absolute configuration: 1R,2R

(assigned by chemical correlation to 2-cyclopenten-1-ol)

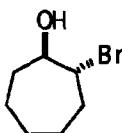
 $C_6H_{11}BrO$ *trans*-2-Bromo-1-cyclohexanol

E.e. >98% [by  $^1\text{H}$ -NMR of the acetate ester in the presence of  $\text{Eu}(\text{hfc})_3$ ]  
 $[\alpha]_D = -27.5$  (*c* 11.2,  $\text{CH}_2\text{Cl}_2$ ), lit.\*  $[\alpha]_D = -33.2$  (*c* 2,  $\text{CH}_2\text{Cl}_2$ )

Source of chirality: enzyme-catalyzed resolution

Absolute configuration: 1R,2R

\*Hönig, H.; Seufert-Wasserthal, P. *Synthesis*, 1990, 12, 1137-1140.

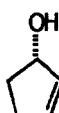


$C_7H_{13}BrO$   
trans-2-Bromo-1-cycloheptanol

E.e. = 99% [by  $^1H$ -NMR of the acetate ester in the presence of Eu(hfc)<sub>3</sub>]  
 $[\alpha]_D = -4.6$  (*c* 10.8,  $CH_2Cl_2$ )

Source of chirality: enzyme-catalyzed resolution

Absolute configuration: 1R,2R  
(assigned by chemical correlation to 2-cyclohepten-1-ol)



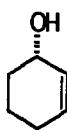
$C_5H_{10}O$   
2-Cyclopenten-1-ol

E.e. = 65%ee [by  $^1H$ -NMR of the Mosher ester]  
 $[\alpha]_D = -91$  (*c* 3.9,  $CCl_4$ ), lit.\* $[\alpha]_D = -106$  (*c* 1.1,  $CHCl_3$ , 82% ee)

Source of chirality: enzyme-catalyzed resolution

Absolute configuration: S

\*Sato, T.; Gotoh, Y.; Wakabayashi, Y.; Fujisawa, T. *Tetrahedron Lett.* 1983, 24, 4123-4126.



$C_6H_{12}O$   
2-Cyclohexen-1-ol

E.e. >99% [by  $^1H$ -NMR of the Mosher ester]  
 $[\alpha]_D = -125$  (*c* 6.4,  $CHCl_3$ ), lit.\* $[\alpha]_D = -97$  (*c* 1.4,  $CHCl_3$ )

Source of chirality: enzyme-catalyzed resolution

Absolute configuration: S

\*Sabol, J. S.; Cregge, R. J. *Tetrahedron Lett.* 1989, 30, 3377-3380.



$C_7H_{14}O$   
2-Cyclohepten-1-ol

E.e. >98% [by  $^1H$ -NMR of the Mosher ester]  
 $[\alpha]_D = -24.9$  (*c* 7.8,  $CH_3OH$ ), lit.\* $[\alpha]_D = -7.5$  (*c* 2,  $CH_3OH$ , <20% ee)

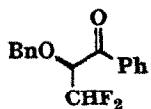
Source of chirality: enzyme-catalyzed resolution

Absolute configuration: S

\*Kasai, M.; Ziffer, H. *J. Org. Chem.* 1983, 48, 712-715.



K. Murata and T. Kitazume

E.e. = >95% [by  $^{19}\text{F}$  NMR with

(+) -MTPA ester]

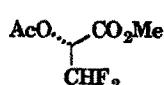
 $[\alpha]_D^{13} +58.78$  ( $c = 0.874$ ,  $\text{CHCl}_3$ )

Absolute configuration S



(S)-(+)-(2-Benzylxyloxy-3,3-difluoro)ethyl phenyl ketone

K. Murata and T. Kitazume

E.e. = >98% [by  $^{19}\text{F}$  NMR with

(+) -MTPA ester]

 $[\alpha]_D^{13} -31.78$  ( $c = 1.10$ ,  $\text{MeOH}$ )

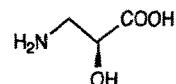
Source of chirality: synthesis

Absolute configuration R (assigned by chem correlation with (R)-(-)-1-phenyl 2,2-difluoroethanol)



(R)-(-)-Methyl acetoxy-3,3-difluorolactate

Y. Lu, C. Miet, N. Kunesch, and J.E. Poisson



E.e. &gt; 99 % [by comparison of optical rotations]

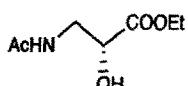
 $[\alpha]_D -32.7$  ( $c = 0.5$ ,  $\text{H}_2\text{O}$ )

m.p. 191-193°C

Source of chirality : Enzymatic kinetic resolution

Absolute configuration : S

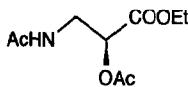
Y. Lu, C. Miet, N. Kunesch, and J.E. Poisson

 $[\alpha]_D -18.7$  ( $c = 3$ ,  $\text{CHCl}_3$ )

Source of chirality : Enzymatic kinetic resolution

Absolute configuration : R

Y. Lu, C. Miet, N. Kunesch, and J.E. Poisson



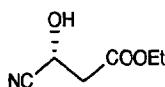
C<sub>9</sub>H<sub>15</sub>NO<sub>5</sub>  
Ethyl *O,N*-Acetyl-3-amino-2-hydroxypropionate

[ $\alpha$ ]<sub>D</sub> +8.5 (c = 3, CHCl<sub>3</sub>)

Source of chirality : Enzymatic kinetic resolution

Absolute configuration : S

Y. Lu, C. Miet, N. Kunesch, and J.E. Poisson



C<sub>6</sub>H<sub>9</sub>NO<sub>3</sub>

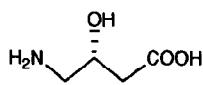
Ethyl 3-Cyano-3-hydroxypropionate

[ $\alpha$ ]<sub>D</sub> +6.7 (c = 2, CHCl<sub>3</sub>)

Source of chirality : Enzymatic kinetic resolution

Absolute configuration : R

Y. Lu, C. Miet, N. Kunesch, and J.E. Poisson



C<sub>4</sub>H<sub>9</sub>NO<sub>3</sub>

gamma-Amino-beta-hydroxybutanoic Acid

E.e. > 99 % [by comparison of optical rotations]

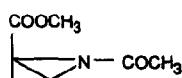
[ $\alpha$ ]<sub>D</sub> -20.9 (c = 1.7, H<sub>2</sub>O)

m.p. 213-215°C

Source of chirality : Enzymatic kinetic resolution

Absolute configuration : R

M. Bucciarelli, A. Forni, I. Monti\*, F. Prati and G. Torre



C<sub>6</sub>H<sub>9</sub>NO<sub>3</sub>

N-Acetyl-2-methoxycarbonylaziridine

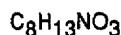
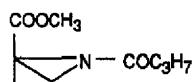
E.e. = 63% [by <sup>1</sup>H nmr with Eu(hfc)<sub>3</sub>]

[ $\alpha$ ]<sub>D</sub><sup>20</sup> = -46.1 (c 1, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration: 2S

(assigned by comparison with literature)



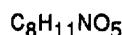
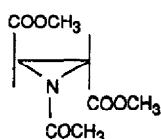
N-Butyryl-2-methoxycarbonylaziridine

E.e. = 90% [by  $^1H$  nmr with Eu(hfc)<sub>3</sub>] $[\alpha]_D^{20} = -74.3$  (c 1, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration: 2S

(assigned by chemical correlation)



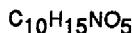
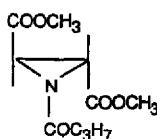
N-Acetyl-2,3-bismethoxycarbonylaziridine

E.e. ≥ 95% [by  $^1H$  nmr with Eu(hfc)<sub>3</sub>] $[\alpha]_D^{20} = -56.4$  (c 1, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration: 2R,3R

(assigned by chemical correlation)



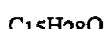
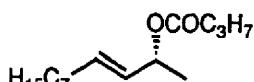
N-Butyryl-2,3-bismethoxycarbonylaziridine

E.e. ≥ 95% [by  $^1H$  nmr with Eu(hfc)<sub>3</sub>] $[\alpha]_D^{20} = -34.8$  (c 1, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration: 2R,3R

(assigned by chemical correlation)



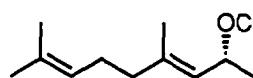
3-Undecen-2-yl Butyrate

E.e. = 91.8% (by GC with acetyl-(S)-lactyl chloride)

Source of Chirality: Enzymatic resolution with PPL

Absolute Configuration: 2R (by GC comparison of acetyl-(S)-lactate esters)

B. Morgan, A.C. Oehlschlager

 $\text{C}_{15}\text{H}_{26}\text{O}_2$ 

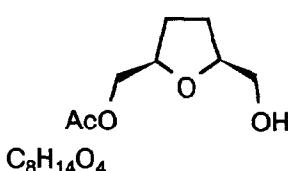
(3E)-4,8-Dimethyl-3,6-nonadien-2-yl Butyrate

E.e. = 95.5% (by GC with acetyl-(S)-lactyl chloride)

Source of Chirality: Enzymatic resolution with PPL

Absolute Configuration: 2R (by GC comparison of acetyl-(S)-lactate esters)

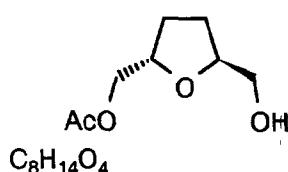
K. Naemura, R. Fukuda, N. Takahashi, M. Konishi, Y. Hirose, and Y. Tobe

 $\text{C}_8\text{H}_{14}\text{O}_4$   
(-)-(2R,5S)-2-(acetoxymethyl)-5-(hydroxymethyl)tetrahydofurane.e. 71% [determined by  $^1\text{H}$  NMR using chiral shift reagent; Eu(hfc)<sub>3</sub>] $[\alpha]_D^{24} -6.17$  (c, 1.10,  $\text{CHCl}_3$ )

Source of Chirality: Enzymatic asymm. hydrolysis and asymm. acylation

Absolute configuration: 2R,5S

K. Naemura, R. Fukuda, N. Takahashi, M. Konishi, Y. Hirose, and Y. Tobe

 $\text{C}_8\text{H}_{14}\text{O}_4$   
(+)-(2S,5S)-2-(acetoxymethyl)-5-(hydroxymethyl)tetrahydofuran

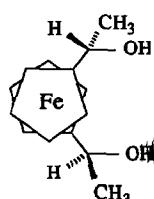
e.e. 58%

 $[\alpha]_D^{24} +22.8$  (c, 1.05,  $\text{CHCl}_3$ )

Source of Chirality: Enzymatic enantioselective hydrolysis and enantioselective acylation

Absolute configuration: 2S,5S

D. Lambusta, G. Nicolosi, A. Patti and M. Piattelli

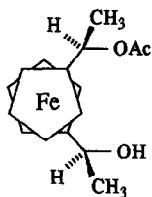
 $\text{C}_{14}\text{H}_{18}\text{FeO}_2$   
(S)-1,1'-bis( $\alpha$ -hydroxyethyl)ferroceneE.e.=100% [by nmr with Eu(hfc)<sub>3</sub>] $[\alpha]_D = +42.0$  (c 0.5,  $\text{C}_6\text{H}_6$ )

Source of chirality: Lipase-mediated esterification

Absolute configuration: S,S

D. Lambusta, G. Nicolosi, A. Patti and M. Piattelli

Tetrahedron: Asymmetry 1993, 4, 919

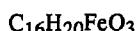


E.e.=90% [by nmr with Eu(hfc)<sub>3</sub>]

$[\alpha]_D = +2.0$  (c 1, CHCl<sub>3</sub>)

Source of chirality: Lipase-mediated esterification

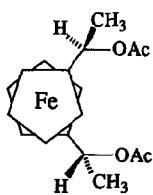
Absolute configuration: R,S



(R)-1-(α-acetoxyethyl)-(S)-1'-(α-hydroxyethyl)ferrocene

D. Lambusta, G. Nicolosi, A. Patti and M. Piattelli

Tetrahedron: Asymmetry 1993, 4, 919

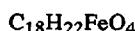


E.e.=100% [by nmr with Eu(hfc)<sub>3</sub>]

$[\alpha]_D = -48.5$  (c 1, CHCl<sub>3</sub>)

Source of chirality: Lipase mediated esterification

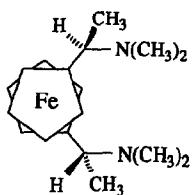
Absolute configuration: R,R



(R)-1,1'-bis(α-acetoxyethyl)ferrocene

D. Lambusta, G. Nicolosi, A. Patti and M. Piattelli

Tetrahedron: Asymmetry 1993, 4, 919



E.e.=100% (by nmr with Pirkle's alcohol)

$[\alpha]_D = +26.8$  (c 1.1, CHCl<sub>3</sub>)

Source of chirality: (R)-1,1'-bis(α-acetoxyethyl)ferrocene

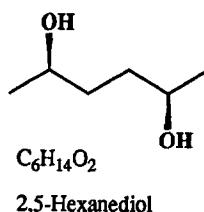
Absolute configuration: R,R



(R)-1,1'-bis[(α-N,N-dimethylamino)ethyl]ferrocene

A. Mattson, N. Örhner, K. Hult, and T. Norin

Tetrahedron: Asymmetry 1993, 4, 925

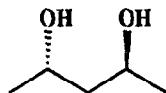


E.e. > 99% (by chiral GC)

D.e. = 98% (by chiral GC)

Source of chirality: resolution by lipase from *Candida antarctica*

Absolute configuration: 2R,5R



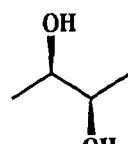
E.e. &gt; 99% (by chiral GC)

D.e. = 94% (by chiral GC)

Source of chirality: resolution by lipase from *Candida antarctica* $C_5H_{12}O_2$ 

2,4-Pentanediol

Absolute configuration: 2S,4S



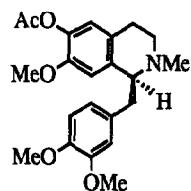
E.e. = 89% (by chiral GC)

D.e. = 61% (by chiral GC)

Source of chirality: resolution by lipase from *Candida antarctica* $C_4H_{10}O_2$ 

2,3-Butanediol

Absolute configuration: 2R,3R



Absolute configuration 1S

E.e. = 96 % on enzymatic resolution  
(estimated by 500 MHz  $^1H$ -NMR analysis of Mosher's ester)[ $\alpha$ ] D + 34.5 (c = 0.49, CHCl<sub>3</sub>) $C_{22}H_{27}NO_5$ 

(S)-(+)-6-Acetoxy-1,2,3,4-tetrahydro-7-methoxy-1-(3,4-dimethoxyphenylmethyl)-2-methylisoquinoline



Absolute configuration 1S

E.e. = 93 % on enzymatic resolution  
(estimated by HPLC analysis using chiralcel OC column)[ $\alpha$ ] D + 25.7 (c = 0.35, CHCl<sub>3</sub>) $C_{31}H_{37}NO_5$ 

(S)-(+)-1,2,3,4-Tetrahydro-7-methoxy-1-(3,4-dimethoxyphenylmethyl)-2-methyl-6-(5-phenylvaleroxy)-isoquinoline

O. Hoshino, R. Tanahashi, M. Okada, H. Akita, T. Oishi

Tetrahedron: Asymmetry 1993, 4, 933



Absolute configuration 1S

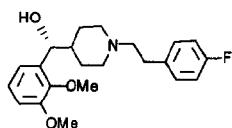
E.e. = 90 % on enzymatic resolution  
(estimated by HPLC analysis of phenol using chiralcel OJ column)

$[\alpha]_D^{25} + 7.5$  ( $c = 1.47$ ,  $\text{CHCl}_3$ )  
 $C_{21}\text{H}_{25}\text{NO}_5$

(S)-(+)-6-Acetoxy-1,2,3,4-tetrahydro-7-methoxy-1-(3,4-dimethoxyphenyl)-2-methylisoquinoline

Chi-Hsin R. King and A.L. Margolin

Tetrahedron: Asymmetry 1993, 4, 943



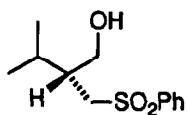
E.e. 96% (Chiralcel OD)  
 $[\alpha]_D^{20} +14.0$  ( $c 0.49$ ,  $\text{CHCl}_3$ )

Source of chirality: lipase-catalyzed resolution

$C_{22}\text{H}_{28}\text{FNO}_3$   
(R)-(+)-4-[1-hydroxy-1-(2,3-dimethoxyphenyl)methyl]-N-2-(4-fluorophenylethyl)piperidine

R. Guevel and L. A. Paquette

Tetrahedron: Asymmetry 1993, 4, 947



$C_{12}\text{H}_{13}\text{O}_3\text{S}$

3-Methyl-2-[(phenylsulfonyl)methyl]-1-butanol

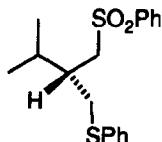
E.e. = 95%

$[\alpha]_D^{20} = -16.3$  ( $c 1.2$ ,  $\text{CHCl}_3$ )

Source of chirality: lipase P30  
hydrolysis of the chloroacetate  
Absolute configuration: S

R. Guevel and L. A. Paquette

Tetrahedron: Asymmetry 1993, 4, 947



$C_{18}\text{H}_{22}\text{O}_2\text{S}_2$

3-Methyl-2-[(phenylsulfonyl)methyl]butyl Phenyl Sulfide

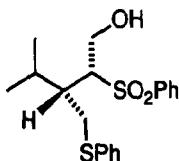
E.e. = 95%

$[\alpha]_D^{20} = -10.1$  ( $c 1.16$ ,  $\text{CHCl}_3$ )

Source of chirality: prepared from enantiomerically enriched alcohol  
Absolute configuration: S

R. Guevel and L. A. Paquette

Tetrahedron: Asymmetry 1993, 4, 947



E.e. = 95%

[α]<sub>D</sub><sup>20</sup> = -140 (c 1.28, CHCl<sub>3</sub>)

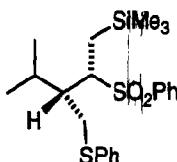
Source of chirality: prepared by alkylation of the enantiomerically enriched sulfone

Absolute configuration: 2R,3S

4-Methyl-2-(phenylsulfonyl)-3-[(phenylthio)methyl]-1-pentanol

R. Guevel and L. A. Paquette

Tetrahedron: Asymmetry 1993, 4, 947



E.e. = 95%

[α]<sub>D</sub><sup>20</sup> = -77 (c 1.34, CHCl<sub>3</sub>)

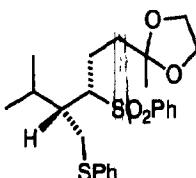
Source of chirality: prepared by alkylation of the enantiomerically enriched sulfone

Absolute configuration: 2R,3S

Trimethyl[4-methyl-2-(phenylsulfonyl)-3-[(phenylthio)methyl]pentyl]silane

R. Guevel and L. A. Paquette

Tetrahedron: Asymmetry 1993, 4, 947



E.e. = 95%

[α]<sub>D</sub><sup>20</sup> = -83 (c 1.26, CHCl<sub>3</sub>)

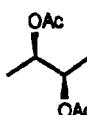
Source of chirality: prepared by alkylation of the enantiomerically enriched sulfone

Absolute configuration: 3S,4S

2-Methyl-2-[5-methyl-3-(phenylsulfonyl)-4-[(phenylthio)methyl]hexyl]-1,3-dioxolane

Kirpal S. Bisht, Virinder S. Panesar and David H.G. Crout

Tetrahedron: Asymmetry 1993, 4, 957



E.e. = >98% [by chiral g.l.c. on a 3-acetyl-2,6-di-O-butyl-β-cyclodextrin column]

[α]<sub>D</sub><sup>20</sup> 14.6 (c, 2.1, CHCl<sub>3</sub>)

Source of chirality: enzyme-catalysed acylation

Absolute configuration 2R, 3R

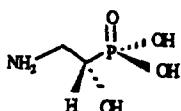
(Assigned by comparison of sign of rotation with compound prepared from authentic 2R, 3R-butanediol)

A. Heisler, C. Rabiller\*, R. Douillard, N. Goalou, G. Hägele and F. Levayer

*Tetrahedron: Asymmetry* 1993, 4, 959

E. e.= 100 % [determined by optical rotation ]

$[\alpha]_D^{22} = +32.5$  (c=1, H<sub>2</sub>O).



2-amino-1-hydroxyethanephosphonic acid

Literature: $[\alpha]_D^{22} = +31.8$  (c=0.525, H<sub>2</sub>O) for the S configuration.

Source of chirality : Pseudomonas Lipase catalysed resolution.

Absolute configuration : S, assigned by comparison with  $[\alpha]_D^{22}$

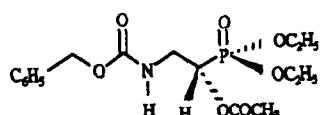
literature data.

A. Heisler, C. Rabiller\*, R. Douillard, N. Goalou, G. Hägele and F. Levayer

*Tetrahedron: Asymmetry* 1993, 4, 959

E. e.= 100 % [determined by optical rotation ]

$[\alpha]_D^{22} = +12.5$  (c=1, CHCl<sub>3</sub>).



Diethyl 1-acetoxy-2-benzyloxycarbonylamino ethanephosphonate.

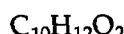
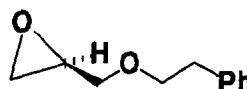
Source of chirality : Pseudomonas Lipase catalysed resolution.

Absolute configuration : S, assigned by comparison of the  $[\alpha]_D^{22}$

value (+32.5, c=1, H<sub>2</sub>O) of the deprotected aminophosphonic acid with literature data: +31.8 (c=0.525, H<sub>2</sub>O) for the S configuration.

V. Partali, V. Waagen, T. Alvik and T. Anthonsen

*Tetrahedron: Asymmetry* 1993, 4, 961



(R)-2-Phenylethyl glycidyl ether

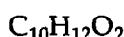
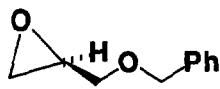
$[\alpha]_D^{20} = -16.7$  (c 1.44, Benzene)

$[\alpha]_D^{20} = -10.5$  (c 1.71, EtOH)

Prepared from (S)-epichlorohydrin and from (S)-1-[2-phenylethyl]-3-chloro-1,2-propanediol

V. Partali, V. Waagen, T. Alvik and T. Anthonsen

*Tetrahedron: Asymmetry* 1993, 4, 961



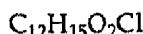
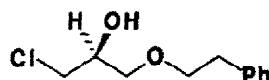
$[\alpha]_D^{20} = -6.2$  (c 1.58, Benzene)

Prepared from (S)-epichlorohydrin and from (S)-1-phenylmethyl-3-chloro-1,2-propanediol

(R)-Phenylmethyl glycidyl ether

V. Partali, V. Waagen, T. Alvik and T. Anthonsen

Tetrahedron: Asymmetry 1993, 4, 961

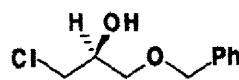


$[\alpha]_D^{20} = +6.5$  (c 1.29, EtOH)  
Prepared from (S)-epichlorohydrin, ee > 99%  
Enzymatic racemate resolution of  
2-butanoate,  $E = 25$  (Amano SAM II)

(S)-1-[2-phenylethyl]-3-chloro-1,2-propanediol

V. Partali, V. Waagen, T. Alvik and T. Anthonsen

Tetrahedron: Asymmetry 1993, 4, 961

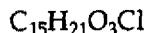
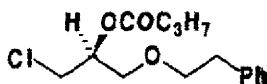


$[\alpha]_D^{20} = +4.5$  (c 2.15, EtOH)  
Prepared from (S)-epichlorohydrin, ee > 99%  
Enzymatic racemate resolution of  
2-butanoate,  $E = 15$  (PPL)

(S)-1-phenylmethyl-3-chloro-1,2-propanediol

V. Partali, V. Waagen, T. Alvik and T. Anthonsen

Tetrahedron: Asymmetry 1993, 4, 961

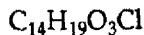
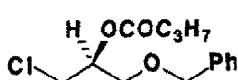


$[\alpha]_D^{20} = +7.1$  (c 2.25, EtOH)  
Prepared from (S)-epichlorohydrin, ee > 99%  
R-Enantiomer from enzymatic  
racemate resolution,  $E = 25$  (Amano SAM II)

(S)-2-butanoyl-1-[2-phenylethyl]-3-chloro-1,2-propanediol

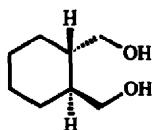
V. Partali, V. Waagen, T. Alvik and T. Anthonsen

Tetrahedron: Asymmetry 1993, 4, 961



$[\alpha]_D^{20} = +10.3$  (c 2.03, EtOH)  
Prepared from (S)-epichlorohydrin, ee > 99%  
R-enantiomer from enzymatic racemate  
resolution,  $E = 15$  (PPL)

(S)-2-butanoyl-1-phenylmethyl-3-chloro-1,2-propanediol



E.e. = 83% (by chiral GC)

 $[\alpha]_D^{25} = -9.48$  ( $c = 1.16, \text{CHCl}_3$ )

Source of chirality: lipase resolution

Absolute configuration: 1S, 2S

SS - *trans* - cyclohexane -1,2 - dimethanolE.e. = >99 % [by  $^1\text{H}$ -NMR of the MTPA ester derivative] $[\alpha]_D^{25} = +7.11$  ( $c = 0.9, \text{CHCl}_3$ )

Source of chirality: Enzymatic hydrolysis

Absolute configuration: R

assigned by chemical correlation

$\text{C}_6\text{H}_{12}\text{O}_3$   
(R)-3-Acetoxy-2-methyl-1-propanol

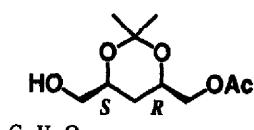


E.e. = &gt;99%

 $[\alpha]_D^{21} = -20.1$  ( $c = 1.6, \text{CHCl}_3$ )

Absolute configuration: S

$\text{C}_5\text{H}_9\text{NBr}$   
(S)-4-Bromo-3-methylbutanenitrile

E.e. = 96 % [by  $^1\text{H}$ -NMR of the MTPA ester derivative] $[\alpha]_D^{25} = -4.6$  ( $c = 1.0, \text{CHCl}_3$ )

Source of chirality: Enzymatic hydrolysis

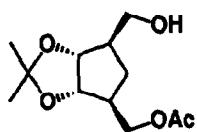
Absolute configuration: 4R,6S

assigned by chemical correlation

$\text{C}_{10}\text{H}_{18}\text{O}_5$   
(4R,6S)-4-Acetoxymethyl-6-hydroxymethyl-2,2-dimethyl-1,3-dioxane

M. Tanaka, M. Yoshioka, and K. Sakai

Tetrahedron: Asymmetry 1993, 4, 981



C<sub>12</sub>H<sub>20</sub>O<sub>5</sub>

(1*R*, 2*R*, 3*S*, 4*S*)-2,3-isopropylidene dihydroxy-cyclopentane-1,4-dimethanol monooacetate

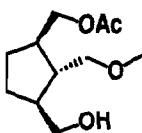
E.e. = >99 % [by <sup>1</sup>H-NMR of the MTPA ester derivative]

[α]<sub>D</sub><sup>22</sup> = -9.17 (c = 1.15, CHCl<sub>3</sub>)

Source of chirality: Enzymatic hydrolysis

M. Tanaka, M. Yoshioka, and K. Sakai

Tetrahedron: Asymmetry 1993, 4, 981



C<sub>11</sub>H<sub>20</sub>O<sub>4</sub>

2-Methoxymethylcyclopentane-1,β-dimethanol monoacetate

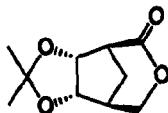
E.e. = >99 % [by <sup>1</sup>H-NMR of the MTPA ester derivative]

[α]<sub>D</sub><sup>25</sup> = -60.60 (c = 0.94, CHCl<sub>3</sub>)

Source of chirality: Enzymatic hydrolysis

M. Tanaka, M. Yoshioka, and K. Sakai

Tetrahedron: Asymmetry 1993, 4, 981



C<sub>10</sub>H<sub>14</sub>O<sub>4</sub>

(1*S*, 5*S*, 6*S*, 7*R*)-6,7-isopropylidene dihydroxy-3-oxabicyclo[3.2.1]octan-2-one

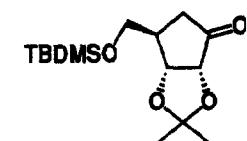
E.e. = >99 %

[α]<sub>D</sub><sup>25</sup> = -41.0 (c = 0.97, CHCl<sub>3</sub>)

Source of chirality: Enzymatic hydrolysis

M. Tanaka, M. Yoshioka, and K. Sakai

Tetrahedron: Asymmetry 1993, 4, 981



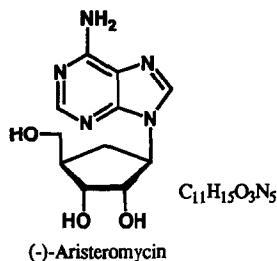
C<sub>15</sub>H<sub>28</sub>O<sub>4</sub>Si

(2*S*, 3*R*, 4*R*)-4-tert-butyl dimethylsilyloxy methyl-2,3-isopropylidene dioxycyclopentanone

E.e. = >99 %

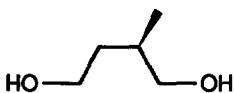
[α]<sub>D</sub><sup>22</sup> = -133.1 (c = 1.00, CHCl<sub>3</sub>)

Source of chirality: Enzymatic hydrolysis



E.e. = >99 %  
 $[\alpha]_D^{25} = -52.1$  ( $c = 0.275$ ,  $\text{CHCl}_3$ )  
 Source of chirality: Enzymatic hydrolysis

Absolute configuration:  
 $(1R, 2S, 3R, 4R)$



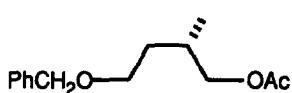
$\text{C}_5\text{H}_{12}\text{O}_2$   
(R)-2-methyl-1,4-butanediol

E.e. = 70%  
(by  $[\alpha]_D$ )  
 $[\alpha]_D +9$  ( $c 1 \text{ CH}_3\text{OH}$ )  
 Source of chirality: *Pseudomonas fluorescens* lipase  
 Absolute configuration: (R)



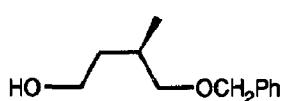
$\text{C}_{12}\text{H}_{18}\text{O}_2$   
(R)-2-methyl-1,4-butanediol,4-benzyl ether

E.e. = 98%  
(by  $^1\text{H-NMR}$  of (R)-MTPA ester)  
 $[\alpha]_D +9.8$  ( $c 4 \text{ C}_2\text{H}_5\text{OH}$ )  
 Source of chirality: *Pseudomonas fluorescens* lipase  
 Absolute configuration: (R)



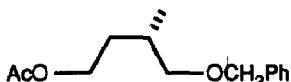
$\text{C}_{14}\text{H}_{20}\text{O}_3$   
(S)-2-methyl-1,4-butanediol,4-benzyl ether,1-acetate

E.e. = 85%  
 $[\alpha]_D +2.4$  ( $c 1 \text{ CH}_3\text{OH}$ )  
 Source of chirality: *Pseudomonas fluorescens* lipase  
 Absolute configuration: (S)



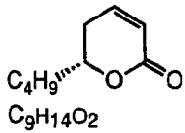
E.e. = 40%  
 (by <sup>1</sup>H-NMR of (R)-MTPA ester)  
 $[\alpha]_D^{20} -0.9$  (c 1 CH<sub>3</sub>OH)  
 Source of chirality: *Pseudomonas fluorescens* lipase  
 Absolute configuration: (R)

C<sub>12</sub>H<sub>18</sub>O<sub>2</sub>  
 (R)-2-methyl-1,4-butanediol,1-benzyl ether



E.e. = 52%  
 $[\alpha]_D^{20} -1.3$  (c 1 CH<sub>3</sub>OH)  
 Source of chirality: *Pseudomonas fluorescens* lipase  
 Absolute configuration: (S)

C<sub>14</sub>H<sub>20</sub>O<sub>3</sub>  
 (S)-2-methyl-1,4-butanediol,1-benzyl ether,4-acetate

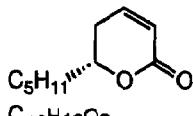


C<sub>4</sub>H<sub>9</sub>  
 C<sub>9</sub>H<sub>14</sub>O<sub>2</sub>

6-Butyl-5,6-dihydro-2H-pyran-2-one

E.e. = > 99% [by GC using Lipodex E]  
 $[\alpha]_D^{20} = -128.8$  (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis  
 Absolute configuration 6R



C<sub>5</sub>H<sub>11</sub>  
 C<sub>10</sub>H<sub>16</sub>O<sub>2</sub>

6-Pentyl-5,6-dihydro-2H-pyran-2-one

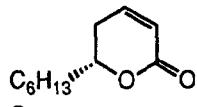
E.e. = > 99% [by GC using Lipodex E]  
 $[\alpha]_D^{20} = -114.5$  (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis  
 Absolute configuration 6R

B. Haase and M. P. Schneider

E.e. = 98% [by GC using Lipodex E]

$[\alpha]_D^{20} = -125.4$  ( $c = 1.0, \text{CHCl}_3$ )



$C_6\text{H}_3$   
 $C_{11}\text{H}_{18}\text{O}_2$

6-Hexyl-5,6-dihydro-2*H*-pyran-2-one

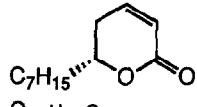
Source of chirality: enzymatic hydrolysis

Absolute configuration 6*R*

B. Haase and M. P. Schneider

E.e. = 96% [by GC using Lipodex E]

$[\alpha]_D^{20} = -78.0$  ( $c = 1.0, \text{CHCl}_3$ )



$C_7\text{H}_{15}$

$C_{12}\text{H}_{20}\text{O}_2$

6-Heptyl-5,6-dihydro-2*H*-pyran-2-one

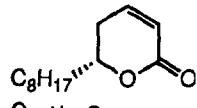
Source of chirality: enzymatic hydrolysis

Absolute configuration 6*R*

B. Haase and M. P. Schneider

E.e. = >99% [by GC using Lipodex E]

$[\alpha]_D^{20} = -86.6$  ( $c = 1.0, \text{CHCl}_3$ )



$C_8\text{H}_{17}$

$C_{13}\text{H}_{22}\text{O}_2$

6-Octyl-5,6-dihydro-2*H*-pyran-2-one

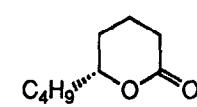
Source of chirality: enzymatic hydrolysis

Absolute configuration 6*R*

B. Haase and M. P. Schneider

E.e. = > 99% [by GC using Lipodex E]

$[\alpha]_D^{20} = +50.6$  ( $c = 1.0, \text{CHCl}_3$ )



$C_4\text{H}_9$

$C_9\text{H}_{16}\text{O}_2$

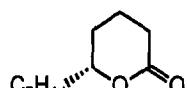
6-Butyl-oxan-2-one

Source of chirality: enzymatic hydrolysis

Absolute configuration 6*R*

B. Haase and M. P. Schneider

Tetrahedron: Asymmetry 1993, 4, 1017



C<sub>5</sub>H<sub>11</sub><sup>...</sup>O  
C<sub>10</sub>H<sub>18</sub>O<sub>2</sub>  
6-Pentyl-oxan-2-one

E.e. = >99% [by GC using Lipodex E]

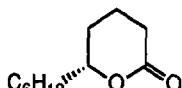
[ $\alpha$ ]<sub>D</sub><sup>20</sup> = +47.2 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 6*R*

B. Haase and M. P. Schneider

Tetrahedron: Asymmetry 1993, 4, 1017



C<sub>6</sub>H<sub>13</sub><sup>...</sup>O  
C<sub>11</sub>H<sub>20</sub>O<sub>2</sub>  
6-Hexyl-oxan-2-one

E.e. = 98% [by GC using Lipodex E]

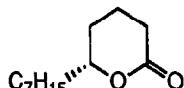
[ $\alpha$ ]<sub>D</sub><sup>20</sup> = +43.7 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 6*R*

B. Haase and M. P. Schneider

Tetrahedron: Asymmetry 1993, 4, 1017



C<sub>7</sub>H<sub>15</sub><sup>...</sup>O  
C<sub>12</sub>H<sub>22</sub>O<sub>2</sub>  
6-Heptyl-oxan-2-one

E.e. = 96% [by GC using Lipodex E]

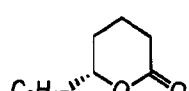
[ $\alpha$ ]<sub>D</sub><sup>20</sup> = +35.3 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 6*R*

B. Haase and M. P. Schneider

Tetrahedron: Asymmetry 1993, 4, 1017



C<sub>8</sub>H<sub>17</sub><sup>...</sup>O  
C<sub>13</sub>H<sub>24</sub>O<sub>2</sub>  
6-Octyl-oxan-2-one

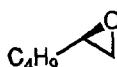
E.e. = >99% [by GC using Lipodex E]

[ $\alpha$ ]<sub>D</sub><sup>20</sup> = +38.4 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 6*R*

B. Haase and M. P. Schneider



C<sub>6</sub>H<sub>12</sub>O  
1,2-Epoxyhexene

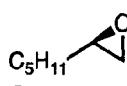
E.e. = 98% [by HPLC as BGIT derivative]

[ $\alpha$ ]<sub>D</sub><sup>20</sup> = +9.1 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 2*R*

B. Haase and M. P. Schneider



C<sub>5</sub>H<sub>10</sub>O  
1,2-Epoxypentene

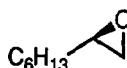
E.e. = >98% [by HPLC as BGIT derivative]

[ $\alpha$ ]<sub>D</sub><sup>20</sup> = +9.8 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 2*R*

B. Haase and M. P. Schneider



C<sub>8</sub>H<sub>16</sub>O  
1,2-Epoxyoctene

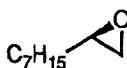
E.e. = 95% [by HPLC as BGIT derivative]

[ $\alpha$ ]<sub>D</sub><sup>20</sup> = +9.8 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 2*R*

B. Haase and M. P. Schneider



C<sub>9</sub>H<sub>18</sub>O  
1,2-Epoxynonene

E.e. = 96% [by HPLC as BGIT derivative]

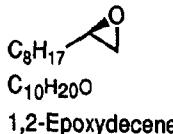
[ $\alpha$ ]<sub>D</sub><sup>20</sup> = +8.1 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 2*R*

B. Haase and M. P. Schneider

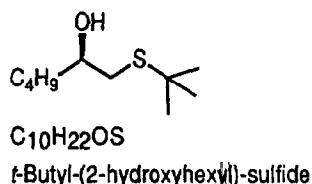
E.e. = 98% [by HPLC as BGIT derivative]  
 $[\alpha]_D^{20} = +7.4$  ( $c = 1.0, \text{CHCl}_3$ )



Source of chirality: enzymatic hydrolysis

Absolute configuration  $2R$

B. Haase and M. P. Schneider

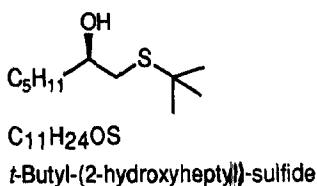


E.e. = > 99% [by GC using Cyclodex  $\beta$  I/P]  
 $[\alpha]_D^{20} = -29.6$ , ( $c = 1.0, \text{CHCl}_3$ )

Source of chirality: enzymatic hydrolysis

Absolute configuration  $R$

B. Haase and M. P. Schneider

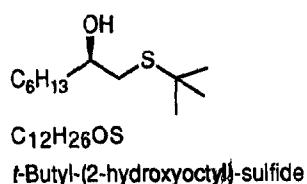


E.e. = > 99% [by GC using Cyclodex  $\beta$  I/P]  
 $[\alpha]_D^{20} = -25.0$ , ( $c = 1.0, \text{CHCl}_3$ )

Source of chirality: enzymatic hydrolysis

Absolute configuration  $R$

B. Haase and M. P. Schneider

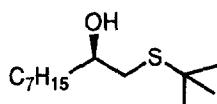


E.e. = 98% [by GC using Cyclodex  $\beta$  I/P]  
 $[\alpha]_D^{20} = -21.5$ , ( $c = 1.0, \text{CHCl}_3$ )

Source of chirality: enzymatic hydrolysis

Absolute configuration  $R$

B. Haase and M. P. Schneider



C<sub>13</sub>H<sub>28</sub>OS  
t-Butyl-(2-hydroxynonyl)-sulfide

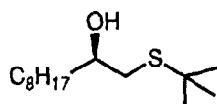
E.e. = 96% [by GC using Cyclodex β I/P]

[α]<sub>D</sub><sup>20</sup> = -20.0, (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration R

B. Haase and M. P. Schneider



C<sub>14</sub>H<sub>30</sub>OS  
t-Butyl-(2-hydroxydecyl)-sulfide

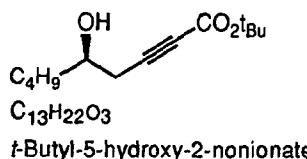
E.e. = >99% [by GC using Cyclodex β I/P]

[α]<sub>D</sub><sup>20</sup> = -19.0, (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration R

B. Haase and M. P. Schneider



C<sub>13</sub>H<sub>22</sub>O<sub>3</sub>  
t-Butyl-5-hydroxy-2-nonionate

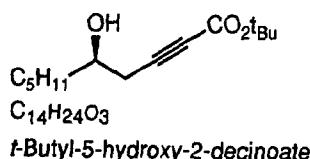
E.e. = > 99% [by precursor]

[α]<sub>D</sub><sup>20</sup> = -6.8 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 5R

B. Haase and M. P. Schneider



C<sub>14</sub>H<sub>24</sub>O<sub>3</sub>  
t-Butyl-5-hydroxy-2-decanoate

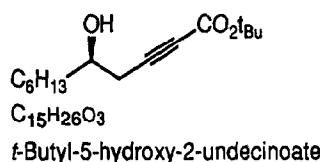
E.e. = > 99% [by precursor]

[α]<sub>D</sub><sup>20</sup> = -6.5 (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

Absolute configuration 5R

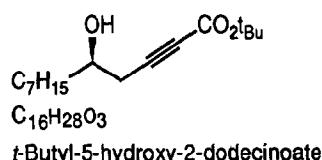
B. Haase and M. P. Schneider



E.e. = 98% [by precursor]  
 $[\alpha]_D^{20} = -3.2$  ( $c = 1.0, \text{CHCl}_3$ )

Source of chirality: enzymatic hydrolysis  
Absolute configuration 5*R*

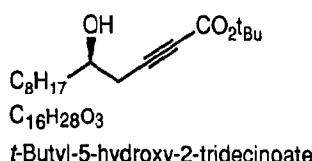
B. Haase and M. P. Schneider



E.e. = 96% [by precursor]  
 $[\alpha]_D^{20} = -5.2$  ( $c = 1.0, \text{CHCl}_3$ )

Source of chirality: enzymatic hydrolysis  
Absolute configuration 5*R*

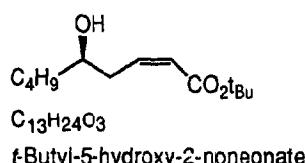
B. Haase and M. P. Schneider



E.e. = >99% [by precursor]  
 $[\alpha]_D^{20} = -5.0$  ( $c = 1.0, \text{CHCl}_3$ )

Source of chirality: enzymatic hydrolysis  
Absolute configuration 5*R*

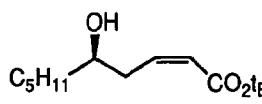
B. Haase and M. P. Schneider



E.e. = > 99% [by precursor]  
 $[\alpha]_D^{20} = +11.1$  ( $c = 1.0, \text{CHCl}_3$ )

Source of chirality: enzymatic hydrolysis  
Absolute configuration 5*R*

B. Haase and M. P. Schneider

 $C_{14}H_{26}O_3$ *t*-Butyl-5-hydroxy-2-decenoate

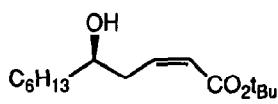
E.e. = &gt; 99% [by precursor]

 $[\alpha]_D^{20} = +9.2$  ( $c = 1.0, CHCl_3$ )

Source of chirality: enzymatic hydrolysis

Absolute configuration 5*R*

B. Haase and M. P. Schneider

 $C_{15}H_{28}O_3$ *t*-Butyl-5-hydroxy-2-undecenoate

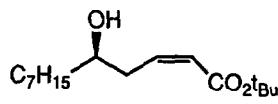
E.e. = 98% [by precursor]

 $[\alpha]_D^{20} = +8.9$  ( $c = 1.0, CHCl_3$ )

Source of chirality: enzymatic hydrolysis

Absolute configuration 5*R*

B. Haase and M. P. Schneider

 $C_{16}H_{30}O_3$ *t*-Butyl-5-hydroxy-2-dodecenoate

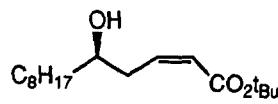
E.e. = 96% [by precursor]

 $[\alpha]_D^{20} = +6.0$  ( $c = 1.0, CHCl_3$ )

Source of chirality: enzymatic hydrolysis

Absolute configuration 5*R*

B. Haase and M. P. Schneider

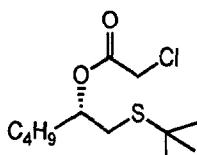
 $C_{17}H_{32}O_3$ *t*-Butyl-5-hydroxy-2-tridecenoate

E.e. = &gt;99% [by precursor]

 $[\alpha]_D^{20} = +7.1$  ( $c = 1.0, CHCl_3$ )

Source of chirality: enzymatic hydrolysis

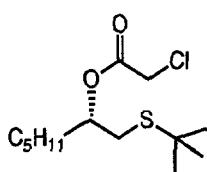
Absolute configuration 5*R*

C<sub>12</sub>H<sub>23</sub>O<sub>2</sub>CIS*t*-Butyl-(2-(chloracetyl)-hexyl)-sulfide

E.e. = > 99% [as *t*-Butyl-(2-hydroxyhexyl)-sulfide]  
 $[\alpha]_D^{20} = -38.7$ , (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

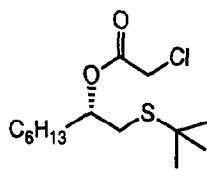
Absolute configuration S

C<sub>13</sub>H<sub>25</sub>O<sub>2</sub>CIS*t*-Butyl-(2-(chloracetyl)-heptyl)-sulfide

E.e. = > 99% [as *t*-Butyl-(2-hydroxyheptyl)-sulfide]  
 $[\alpha]_D^{20} = -35.2$ , (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

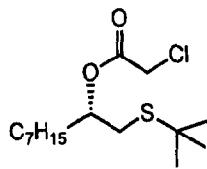
Absolute configuration S

C<sub>14</sub>H<sub>27</sub>O<sub>2</sub>CIS*t*-Butyl-(2-(chloracetyl)-octyl)-sulfide

E.e. = 98% [as *t*-Butyl-(2-hydroxyoctyl)-sulfide]  
 $[\alpha]_D^{20} = -34.9$ , (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

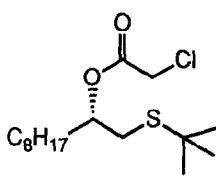
Absolute configuration S

C<sub>15</sub>H<sub>29</sub>O<sub>2</sub>CIS*t*-Butyl-(2-(chloracetyl)-nonyl)-sulfide

E.e. = >99% [as *t*-Butyl-(2-hydroxynonyl)-sulfide]  
 $[\alpha]_D^{20} = -30.1$ , (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis

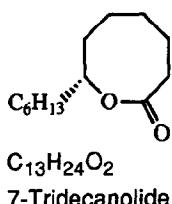
Absolute configuration S



C<sub>16</sub>H<sub>30</sub>O<sub>2</sub>CIS  
t-Butyl-(2-(chloracetyl)-decyl)-sulfide

E.e. = >99% [as *t*-Butyl-(2-hydroxydecyl)-sulfide]  
[ $\alpha$ ]<sub>D</sub><sup>20</sup> = -28.0, (c = 1.0, CHCl<sub>3</sub>)

Source of chirality: enzymatic hydrolysis  
Absolute configuration *S*

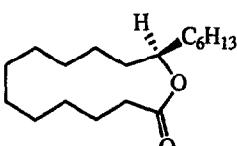


C<sub>13</sub>H<sub>24</sub>O<sub>2</sub>  
7-Tridecanolide

[ $\alpha$ ]<sub>D</sub><sup>20</sup> = -2.4 (c = 0.5, CHCl<sub>3</sub>)

Source of chirality: enzymatic, irreversible  
acyltransfer

Absolute configuration *R*

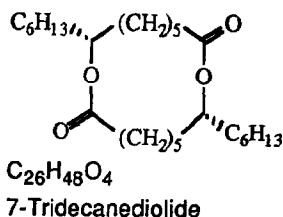


C<sub>18</sub>H<sub>34</sub>O<sub>2</sub>  
12-Octadecanolide

E.e.: > 99 % (specific rotation)  
[ $\alpha$ ]<sub>D</sub><sup>20</sup> = -5.5 (c = 1, CHCl<sub>3</sub>)

Source of chirality: enzymatic, irreversible  
acyltransfer

Absolute configuration *R*  
(assigned to Lit.<sup>3</sup>)

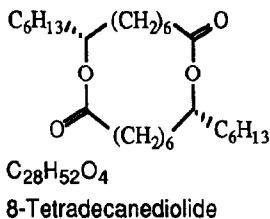


C<sub>26</sub>H<sub>48</sub>O<sub>4</sub>  
7-Tridecanediolide

[ $\alpha$ ]<sub>D</sub><sup>20</sup> = -6.2 (c = 0.5, CHCl<sub>3</sub>)

Source of chirality: enzymatic, irreversible  
acyltransfer

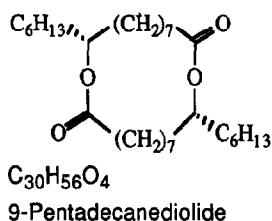
Absolute configuration *R,R*



$[\alpha]_D^{20} = -3.8$  ( $c = 1, \text{CHCl}_3$ )

Source of chirality: enzymatic, irreversible acyltransfer

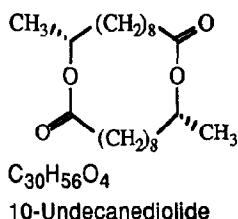
Absolute configuration *R,R*



$[\alpha]_D^{20} = -7.9$  ( $c = 0.9, \text{CHCl}_3$ )

Source of chirality: enzymatic, irreversible acyltransfer

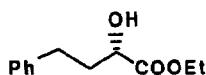
Absolute configuration *R,R*



$[\alpha]_D^{20} = -4.7$  ( $c = 1.7, \text{CHCl}_3$ )

Source of chirality: enzymatic, irreversible acyltransfer

Absolute configuration *R,R*

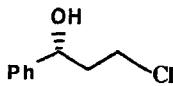


E.e. 94% by comparison on HPLC on chiral column  
Source of chirality: resolution with immobilized Pen-G acylase  
Absolute configuration: *S*



2-Hydroxy-4-phenyl-ethylbutanoate

E. Baldaro, P. D'Arrigo, G. Pedrocchi-Fantoni, C.M. Rosell, S. Servi  
A. Tagliani and M. Terreni



E.e. >98% by comparison on GC on chiral column  
Source of chirality: resolution with immobilized Pen-G acylase  
Absolute configuration: *R*



1-Phenyl-3-chloro-propanol

E. Baldaro, P. D'Arrigo, G. Pedrocchi-Fantoni, C.M. Rosell, S. Servi  
A. Tagliani and M. Terreni

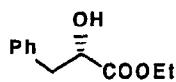


E.e. 40% by comparison on HPLC on chiral column  
Source of chirality: resolution with immobilized Pen-G acylase  
Absolute configuration: *R*



2-Phenyl-ethanol

E. Baldaro, P. D'Arrigo, G. Pedrocchi-Fantoni, C.M. Rosell, S. Servi  
A. Tagliani and M. Terreni

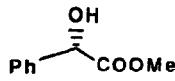


E.e. 92% by comparison on GC on chiral column  
Source of chirality: resolution with immobilized Pen-G acylase  
Absolute configuration: *S*



2-Hydroxy-3-phenyl-ethylpropanoate

E. Baldaro, P. D'Arrigo, G. Pedrocchi-Fantoni, C.M. Rosell, S. Servi  
A. Tagliani and M. Terreni



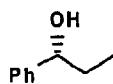
E.e. 90% by comparison on HPLC on chiral column  
Source of chirality: resolution with immobilized Pen-G acylase  
Absolute configuration: *S*



Methyl mandelic acid ester

E. Baldaro, P. D'Arrigo, G. Pedrocchi-Fantoni, C.M. Rosell, S. Servi  
A. Tagliani and M. Terreni

Tetrahedron: Asymmetry 1993, 4, 1031



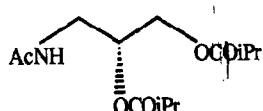
E.e. 94% by comparison on GC on chiral column  
Source of chirality: resolution with immobilized Pen-G acylase  
Absolute configuration: R



1-Phenyl-propanol

M.-A. Mbappé and S. Sicsic

Tetrahedron: Asymmetry 1993, 4, 1035



3-(acetylamino)-1,2-propanediol diisobutyrate

ee=83%

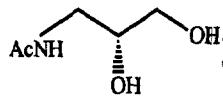
Liquid chromatography of dioxolanes

Source of chirality: resolution (enzym.)

Absolute configuration: R  
(optical rotation of deacylated derivative)

M.-A. Mbappé and S. Sicsic

Tetrahedron: Asymmetry 1993, 4, 1035



ee=67%

Liquid chromatography of dioxolanes

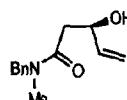
Source of chirality: resolution (enzym.)

Absolute configuration: R

3-(acetylamino)-1,2-propanediol

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose

Tetrahedron: Asymmetry 1993, 4, 1041

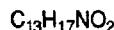


E.e.= > 99% (by HPLC with chiral column)

[α]<sub>D</sub><sup>24</sup>= +24.27 (c 1.425, CHCl<sub>3</sub>)

Source of chirality: enzymatic kinetic resolution

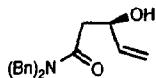
Absolute configuration 3R



(assigned by conversion to the known compound)

N-benzyl-N-methyl-3-hydroxy-4-pentenamide

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose



E.e.= &gt; 99% (by HPLC with chiral column)

 $[\alpha]_D^{25} = +10.73$  (c 1.05, CHCl<sub>3</sub>)

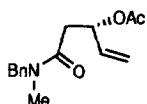
Source of chirality: enzymatic kinetic resolution

Absolute configuration 3R

(assigned by conversion to the known compound)

*N,N*-dibenzyl-3-hydroxy-4-pentenamide

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose



E.e.= 98% (by HPLC with chiral column)

 $[\alpha]_D^{24} = +19.77$  (c 1.825, CHCl<sub>3</sub>)

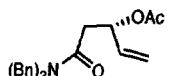
Source of chirality: enzymatic transesterification

Absolute configuration 3S

(assigned by conversion to the known compound)

*N*-benzyl-*N*-methyl-3-acetoxy-4-pentenamide

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose



E.e.= &gt; 99% (by HPLC with chiral column)

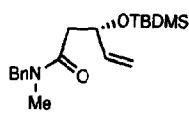
 $[\alpha]_D^{25} = +24.74$  (c 0.88, CHCl<sub>3</sub>)

Source of chirality: enzymatic transesterification

Absolute configuration 3S

*N,N*-dibenzyl-3-acetoxy-4-pentenamide*N,N*-dibenzyl-3-acetoxy-4-pentenamide

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose



E.e.= 97% (by HPLC with chiral column)

 $[\alpha]_D^{25} = -6.36$  (c 2.92, CHCl<sub>3</sub>)

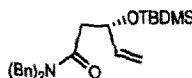
Source of chirality: prepared from homochiral alcohol

Absolute configuration 3S

*C*<sub>19</sub>*H*<sub>31</sub>*NO*<sub>2</sub>*Si**N*-benzyl-*N*-methyl-3-*tert*-butyldimethylsilyloxy-4-pentenamide

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose

Tetrahedron: Asymmetry 1993, 4, 1041

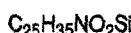


E.e.= 97% (by HPLC with chiral column)

$[\alpha]_D^{25} = -5.67$  (c 3.835, CHCl<sub>3</sub>)

Source of chirality: prepared from homochiral alcohol

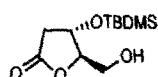
Absolute configuration 3S



N,N-dibenzyl-3-tert-butylidimethylsilyloxy-4-pentenamide

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose

Tetrahedron: Asymmetry 1993, 4, 1041

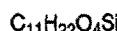


E.e.= 99% (by HPLC with chiral column)

$[\alpha]_D^{25} = +37.33$  (c 3.405, CHCl<sub>3</sub>)

Source of chirality: prepared from homochiral amide

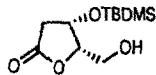
Absolute configuration 3S, 4R



3-O-tert-butylidimethylsilyl-2-deoxy-D-ribo-1,4-lactone

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose

Tetrahedron: Asymmetry 1993, 4, 1041

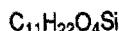


E.e.= 99% (by HPLC with chiral column)

$[\alpha]_D^{25} = -8.73$  (c 1.805, CHCl<sub>3</sub>)

Source of chirality: enzymatic transesterification

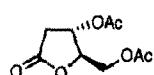
Absolute configuration 3S, 4S



3-O-tert-butylidimethylsilyl-2-deoxy-L-xylo-1,4-lactone

H. Takahata, Y. Uchida, Y. Ohkawa, and T. Momose

Tetrahedron: Asymmetry 1993, 4, 1041



E.e.= 99% (by HPLC with chiral column)

$[\alpha]_D^{25} = -5.31$  (c, 1.25, EtOH)

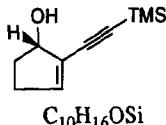
Source of chirality: prepared from homochiral lactone

Absolute configuration 3S, 4R



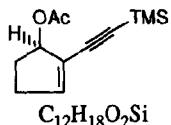
di-O-acetyl-2-deoxy-D-ribo-1,4-lactone

Seiichi Takano,\* Mahito Suzuki, and Kunio Ogasawara



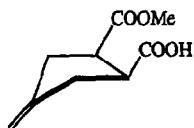
Absolute configuration S  
mp 43.0 °C  
[α]<sub>D</sub><sup>30</sup> -19.8 (c 0.59, CHCl<sub>3</sub>)  
source of chirality: enzymatic transesterification  
E. e.=≥99% by chiral HPLC

Seiichi Takano,\* Mahito Suzuki, and Kunio Ogasawara



Absolute configuration S  
[α]<sub>D</sub><sup>29</sup> -53.6 (c 1.08, CHCl<sub>3</sub>)  
source of chirality: enzymatic transesterification  
E. e.=≥99% by chiral HPLC

Peter Renold and Christoph Tamm



C<sub>9</sub>H<sub>12</sub>O<sub>4</sub>

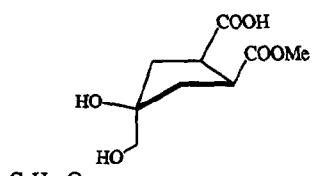
2-(Methoxycarbonyl)-4-methylen-cyclopentane-1-carboxylic acid

E. e.= 63% [ GC, derivative with (1S)-Phenylethyl amine]

Source of chirality: enzymatic hydrolysis

Absolute configuration: 1S, 2R

Peter Renold and Christoph Tamm



2-(Methoxycarbonyl)-4-hydroxy-4-hydroxymethyl-cyclopentane-1-carboxylic acid

E. e.= 73% [ GC, derivative with (1S)-Phenylethyl amine]

Source of chirality: enzymatic hydrolysis

Absolute configuration: 1R, 2S, 4S

Peter Renold and Christoph Tamm



E. e.= 60% [ GC on chiral column of the esters]

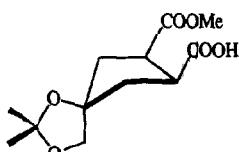
Source of chirality: enzymatic hydrolysis

Absolute configuration: 1R, 4S, 5R

 $C_8H_{10}O_5$ 

1-Hydroxymethyl-3-oxo-2-oxaspiro[3.1.1]heptane-5-carboxylic acid

Peter Renold and Christoph Tamm



E. e.= 64% [ GC, derivative with (1S)-Phenylethyl amine]

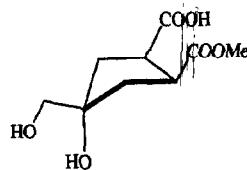
Source of chirality: enzymatic hydrolysis

Absolute configuration: 5R, 7S, 8R

 $C_{12}H_{18}O_6$ 

8-(Methoxycarbonyl)-2,2-dimethyl-1,3-dioxaspiro[4.4]nonane-7-carboxylic acid

Peter Renold and Christoph Tamm



E. e.= 44% [ GC, derivative with (1S)-Phenylethyl amine]

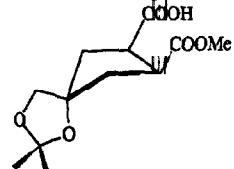
Source of chirality: enzymatic hydrolysis

Absolute configuration: 1R, 2S, 4R

 $C_9H_{14}O_6$ 

2-(Methoxycarbonyl)-4-hydroxy-4-hydroxymethyl-cyclopentane-1-carboxylic acid

Peter Renold and Christoph Tamm



E. e.= 6% [ GC, derivative with (1S)-Phenylethyl amine]

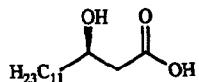
Source of chirality: enzymatic hydrolysis

Absolute configuration: 5S, 7R, 8S

 $C_{12}H_{18}O_6$ 

8-(Methoxycarbonyl)-2,2-dimethyl-1,3-dioxaspiro[4.4]nonane-7-carboxylic acid

T. Sugai, H. Ritzén and C.-H. Wong



$\text{C}_{14}\text{H}_{28}\text{O}_3$   
3-(*R*)-hydroxytetradecanoic acid

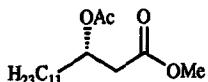
E.e. = 98% (by nmr with MTPA-ester)

 $[\alpha]_D^{25} = 15.1$  (*c* 1.1,  $\text{CHCl}_3$ )

M.p. = 72.0-72.5°C

Source of chirality: Enzymatic resolution

T. Sugai, H. Ritzén and C.-H. Wong



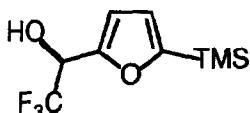
$\text{C}_{17}\text{H}_{32}\text{O}_4$   
3-(*S*)-acetoxytetradecanoic acid

E.e. = 70% (by nmr with MTPA-ester of the hydroxyester)

 $[\alpha]_D^{25} = -1.3$  (*c* 3.1,  $\text{CHCl}_3$ )

Source of chirality: Enzymatic resolution

T. Yamazaki, K. Mizutani, and T. Kitazume

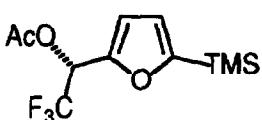


$\text{C}_9\text{H}_{13}\text{F}_3\text{O}_2\text{Si}$

(1'*S*)-2-[1'-(2',2',2'-Trifluoro-1'-hydroxyethyl)]-5-trimethylsilylfuranE.e. = 98% [by  $^1\text{H}$  NMR analysis of its MTPA ester] $[\alpha]_D^{27} +7.45$  (*c* 1.10, MeOH)Absolute configuration : *S* [chemical correlation into the reported optically active trifluorinated lactate]

Source of Chirality : Enzymatic optical resolution

T. Yamazaki, K. Mizutani, and T. Kitazume

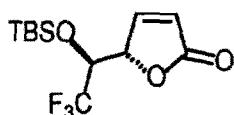


$\text{C}_{11}\text{H}_{15}\text{F}_3\text{O}_3\text{Si}$

(1'*R*)-2-[1'-(1'-Acetoxy-2',2',2'-trifluoroethyl)]-5-trimethylsilylfuranE.e. = 94% [by  $^1\text{H}$  NMR analysis of the MTPA ester after hydrolysis] $[\alpha]_D^{29} -102.68$  (*c* 1.28, MeOH)Absolute configuration : *R*

Source of Chirality : Enzymatic optical resolution

T. Yamazaki, K. Mizutani, and T. Kitazume



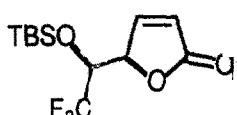
E.e. = 98%

 $[\alpha]_D^{27} -102.42$  (*c* 1.00, MeOH)

Absolute configuration : 1'S,4S [chemical correlation into the reported tris TBS ether after hydrogenation and reduction]

Relative configuration : *anti* $C_{12}H_{19}F_3O_3Si$ (1'S,4S)-4-[1'-(1'-*t*-Butyldimethylsiloxy-2',2',2'-trifluoroethyl)]-2-buten-4-olate

T. Yamazaki, K. Mizutani, and T. Kitazume



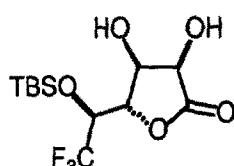
E.e. = 98%

 $[\alpha]_D^{27} +87.86$  (*c* 0.66, MeOH)

Absolute configuration : 1'S,4R [chemical correlation into the reported tris TBS ether after hydrogenation and reduction]

Relative configuration : *syn* $C_{12}H_{19}F_3O_3Si$ (1'S,4R)-4-[1'-(1'-*t*-Butyldimethylsiloxy-2',2',2'-trifluoroethyl)]-2-buten-4-olate

T. Yamazaki, K. Mizutani, and T. Kitazume



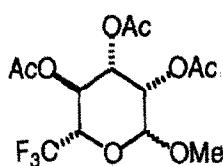
E.e. = 98%

 $[\alpha]_D^{25} +31.20$  (*c* 0.67, CHCl<sub>3</sub>)

Absolute configuration : 1'S,3R,4S,5S [from mechanistic consideration]

 $C_{12}H_{21}F_3O_5Si$ (1'S,3R,4S,5S)-5-[1'-(1'-*t*-Butyldimethylsiloxy-2',2',2'-trifluoroethyl)]-3,4-dihydroxydi-hydro-2(3H)-furanone

T. Yamazaki, K. Mizutani, and T. Kitazume



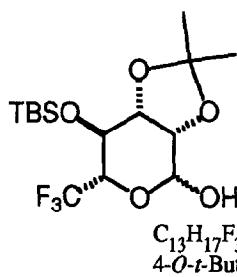
E.e. = 98%

 $[\alpha]_D^{20} +52.69$  (*c* 0.82, CHCl<sub>3</sub>)

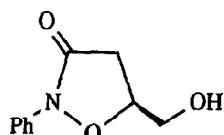
Absolute configuration : 2S,3S,4R,5S [from mechanistic consideration]

 $C_{13}H_{17}F_3O_8$ 

Methyl 2,3,4-O-triAcetyl-6-deoxy-6,6,6-trifluoro-D-manno-hexopyranoside

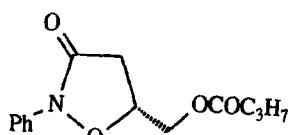


E.e. = 98%  
 $[\alpha]_D^{17} +13.50 (c\ 0.83, \text{CHCl}_3)$   
 Absolute configuration :  $2S,3S,4R,5S$  [from mechanistic consideration]



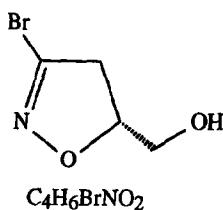
(S)-2-Phenyl-5-hydroxymethyl-isoxazolidin-3-one  
 $[\alpha]_{20}^D +12.33 (c1.01, \text{MeOH})$   
 E.e.=91%

HPLC Chiralcel OB

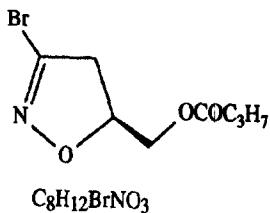


(R)-2-Phenyl-5-hydroxymethyl-isoxazolidin-3-one butyrate  
 $[\alpha]_{20}^D -19.19 (c0.964, \text{MeOH})$   
 E.e.=98%

HPLC Chiralcel OB

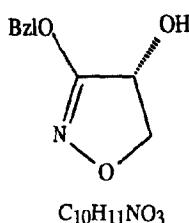


(R)-3-Bromo-5-hydroxymethyl-Delta^2-isoxazoline  
 $[\alpha]_{20}^D -130.25 (c1.01, \text{CHCl}_3)$   
 E.e.=94% HPLC Chiralcel OB

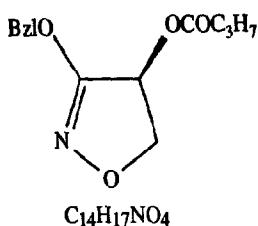


(S)-3-Bromo-5-hydroxymethyl-  
 $\Delta^2$ -isoxazoline butyrate  
 $[\alpha]_{20}^D +98.79(c0.992, \text{CHCl}_3)$   
 E.e. >99%

HPLC Chiralcel OB

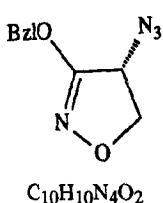


(R)-3-Benzylxy-4-hydroxy-  
 $\Delta^2$ -isoxazoline  
 $[\alpha]_{20}^D +72.46(c1.1, \text{CHCl}_3)$   
 E.e. >99% HPLC Chiralcel OJ



(S)-3-Benzylxy-4-hydroxy-  
 $\Delta^2$ -isoxazoline butyrate  
 $[\alpha]_{20}^D -32.65(c1.07, \text{CHCl}_3)$   
 E.e. >99%

HPLC Chiralcel OJ



(R)-3-Benzylxy-4-azido- $\Delta^2$ -isoxazoline  
 E.e. =99% by HPLC (Chiralcel OJ)  
 $[\alpha]_{20}^D +194.20(c1.0, \text{CHCl}_3)$

Source of chirality: enzymatic resolution

Absolute configurations: chemical correlation with chiral acetyl cycloserines.